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Agriculture

Elixir Agriculture 30 (2011) 1837-1842

Review on infection biology of uromyces species and other rust spores

Sharad Shroff, Dewprakash Patel and Jayant Sahu Banaras Hindu University Varanasi-221005(India).

ARTICLE INFO

Article history: Received: 8 January 2011; Received in revised form: 26 January 2011 Accepted: 29 January 2011

Keywords

penetration hypha, Host surface penetration, Aecium cup, Peridium layer.

ABSTRACT

Uromyces fabae (Uromyces viciae-fabae) the pea rust was first reported by D. C. H. Persoon in 1801. Later DeBary (1862) changed the genus and renamed it as *Uromyces fabae* (Pers) deBary. There after, Kispatic (1949) described *f. sp. viciae-fabae* by including host *vicia fabae*. The pathogen *Uromyces fabae* described as autoecious rust with aeciospores, urediospores and teliospores found on the surface of host plant (Arthur and Cummins, 1962; Gaumann, 1998). Gaumann proposed that the fungus be classified into nine *forma speciales* each with a host range limited to two or there species. Later it was observed that the isolates of *Uromyces viciae-fabae* share so many hosts in common that it was impossible to classify them *into forma speciales* (Conner and Bernier, 1982). Based on the distinctive shape and dimensions of substomatal vesicle, *Uromyces viciae fabae* has been described as a species complex (Emeran *et al.*, 2005). It revealed that host specialized isolates of *Uromyces viciae fabae* were morphologically distinct, differing in both spore dimensions and infection structure.

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Introduction

Uppal (1933) and Prasada and Verma (1948) found that several species of *Vicia*, *Lathyrus*, *Pisum*, and Lentil are susceptible to Uromyces *fabae* in India and abroad. In India species of Vicia, Lathyrus and Pisum are described as host plant for *Uromyces fabae* (pers) deBary, (Kapooria and Sinha, 1966; Kapooria and Sinha, 1971).

They recorded natural infection on *Vicia sativa* L. and *V. hirsuta* Gray, common weeds found in the field of lentil in India also. Viciae faba L., V. biennes L., V. *hirsuta*, and V. arborensis *L.* were described as highly susceptible to *Uromyces fabae*, *Viciae sativa* and *Lathyrus* aphaca were found to be disease free. Conner and Bernier (1982) reported a total of 52 species of *Viciae fabae* and 22 species of *Lathyrus* to be infected by Uromyces viciae *fabae*. They also found this pathogen on pea, lentil and *fababean*. Butler (1912) reported the occurrence of rust pathogen (*Uromyces fabae*) on pea and other leguminous crops from India.

Distribution

There were reports on occurrence of Uromyces fabae from most of the place in India. Butler (1912) reported this fungus from state of Maharastra. Pea rust (Uromyces fabae) is of world wide occurrence and attacks number of host species belonging to different genera of family leguminoseae in the Indo-gangetic plains Butler (1918) and Prasada & Verma (1948) also reported the occurrence of Uromyces fabae on lentil crops from Delhi. Roy (1949) in his list of fungi of Bengal recorded the prevalence of Uromyces fabae on the leaves and stems of *Pisum sativum*. Mitter and Tandon (1930), Patel (1943), Pavgi and Upadhyay (1966) and Kapooria and Sinha (1966) reported the distribution of this pathogen in the regions of Uttar Pradesh. Bilgrami et al., (1979) reported the occurrence of this pathogen on various host species of pea, lathyrus and lentil. Baruah et al., (1979) reported on the pea plants caused by both U. fabae and U. pisi is of rare occurrence in India. Occurence of U. fabae is also availablethe rus from Canada (Conner and Bernier, 1982; Xue and Warkentin, 2002).

Variability in the pathogen

Pathogenic variability has been reported in field collection of *Uromyces fabae* (Singh and Sokhi, 1980; Conner and Bernier, 1982; Xue and Warkentin, 2002). The urediospores of *Uromyces fabae* were the infective spore and used in various resistance screening programme in pea (Xue and Warkentin, 2002); faba bean (Sillero *et al.*, 2000); lentil (Chauhan *et al.*, 1966) and sweet pea (Sokhi, 1984).

Mode of survival of the pathogen.

The aeciospores and urediospores of *Uromyces fabae* did not survive at temperature more then 30°C for one week; therefore, they were not supposed to survive at high temperature of the intervening crop season. Teliospore survives the intervening season and germinates to produce basidiospores Pycnia, developed from growth of basidiospores subsequently caused infection in pea (Singh, 1999). Conner and Bernier (1982) have suggested the role of collateral host in the recurrence of disease. They reported that *Vicia* species and *Lathyrus* species served as a collateral host to *Uromyces fabae* and helped in its survival during the absence of the main crop. **Infection process of rust fungi**

Attachment of rust spores in host surface

Deising *et al.*, (1992) observed that the urediospores of rust fungus form adhesion pad release a cutinase and two specific esterases after contacting the host cuticle, apparently adhesion of the pad is improved by these enzymes Cutinase/esterase inhibitors altered this pattern of adhesion.

Hickman and Epstein (1988) has been described adhesion of fungal propagules to plant or artificial modal surface. Nicholson and Epstein (1991) analyzed chemical on polysaccharides, protein or glycoprotein. Beckett *et al.*, (1990) and Read *et al.*, (1992) found germtube of *Puccinia hordei* when fractured away from the barley cuticles, wax crystals adhered tightly to be underside of the germtube.

Germination of aeciospores

Hiratsuku and Powell (1966) studied the germinating aeciospores in case of *P. harknessii* and *P. stalactiforme* and





Found that binucleate spores of *P. stalactiforme* gave rises to nonseptum germtube with two nuclei. Meinecke, (1929) andNighswander (1963) have noted that binucleate spores produced hyphae had one nucleus per cell. A similar observation was made by True (1939) for the woodgate rust fungus *P. harknessii*. Joseph and Hering (1996) reported that urediospores of *U. viciae-fabae* (Broad bean rust) germinated well in the range 5-26 $^{\circ}$ C, fastest germination at 20 $^{\circ}$ C exposure to 30 $^{\circ}$ C gave poor germination and damaged the spores. Joseph and Hering (1996) also reported that at 20 $^{\circ}$ C some infection occurred within 4 hours leaf wetness, but longer wetness periods up to 24 hours gave increased infection. Relative humidity of 100% was favorable for aeciospores germination while 98% RH favoured urediospores germination 2% was observed at 25 $^{\circ}$ C.

Appresorium formation

Maheswari et al., (1967) reported that heat shock induced the formation of appresorium. Davies and Butler (1968) and Mendgen (1973) reported that appressoria attach firmly to their substrates by means of extracellular adhesives or above stomatal pores appressoria of many rust fungi get by wedging their base between the outer edges of the stomatal opening. Macko et al., (1978) and Grumbow (1977) found chemical stimuli include metabolites extracted from urediospores, for appesoria formation. Kaminsky and Day (1984) reported that ions like K⁺⁺ and Ca⁺⁺, sucrose also acts stimuli for appresorium development. Hoch and Staples (1984) reported that cyclic nucleotides or stimulators of adenylates cyclase also responsible for appresorium formation. Gold and Mendgen (1984) observed that basidiospores of Uromyces appendiculatus penetrates mostly from appressoria, but some times penetrates directly with very short germtube. Kapooria and Mendgen (1985) reported that the uredosporelings of U. appendiculatus, U. viciae-fabae, Phakopsora packyrhizi and aeciosporelings of Arthuriomyces peckianus, nuclear division was strictly correlated with appresorium development including septum formation. Hoch et al., (1987) reported that topographical signals such as scratches or precisely defined ridges on a membrane also induce the formation of appresorium. Bourette and Howard (1990) reported that fungi recognized physical differences in topography and rigidity of the substrates before appressoria formed. Kwon and Hoch (1991) reported that the germtube of U. appendiculatus senses inductive topographies such as stomatal opening or a precisely defined ridges of 0.5 µm height over which the appresorium will be positioned within 4 minutes of initial contact. Bourette and Howard (1990), Kwon and Hoch (1991) reported U. appendiculatus and M. grisea need only 40-50 minutes to form a septum that delinates the appresorium. Read et al., (1992) reported that the mechanism by which a hyphal tip could sense minute surface features may includes the different components of the cytoskeleton or an ionic or electric changes mediated by mechanosensitive channels. Bourette and Howard (1991) and Kwon et al., (1991) also reported that the substrate, microtubules and microfilaments were arranged in a reticulate pattern, close to appressorial wall, with peripheral plaques adjacent to the plasma membrane. Deising et al., (1991) reported that appressoria detach easily from their substrates if they are produced on artificial membranes because the infection peg was able to lift the appresorium from the substrates. Mims and Dyke (1991) reported that appressoria of uredosporelings from rust fungi penetrate through stomata. Swann and Mims (1991) also studied that there were many fungi that penetrate the epidermis directly without producing appressorial cones. Chand and

Kushwaha (2006) observed that only about 1% of germling formed appresorium, but most of the appresorium were found away from stomata.

Penetration hypha

Chang *et al.*, (1981) reported that monokaryotic hypha of rust fungi was constricted during penetration, but fungal wall did not exhibit obvious specialization during penetration of the host wall. Koch *et al.*, (1983) reported that rust fungus in dikaryotic stage, developed appressoria and a penetration hypha. Gold and Mendgen (1984), Mims and Richard (1989) reported monokaryotic, appressoria were basely developed and the penetration hypha was specialized. Harder and Chang (1991) found very specialized haustoria mother cell during intercellular growth.

Host surface penetration

Xu and Mendgen (1991) found that basidiospores of U. vignae can penetrate the host cuticle of Vicia faba much faster and with higher efficiency then the host epidermis, but the infection was stopped a few hours later within the cytoplasm. Heiler et al., (1993) reported that the uredosporelings of Uromyces viciae-fabae produced a cellulolytic enzyme that was regulated in a strickly differentiation in specific manners. Heiler et al., (1993) reported that production of these enzymes continued during later stages of infection hypha when haustorial mother cells were formed.

Infection hypha and differentiation of hypha

Hoch and Staples (1983) observed that the endoplasmic reticulum appears more differentiated during growth (within the host plant) of germtube of *Uromyces appendiculatus*. Gold and Mendgen (1984) found that monokaryotic infections, by basidiospores of rust fungi produce a short germtube, modestly developed appresorium and a penetration hypha which elongates to form intraepidermal vesicles with a primary hypha.

Gold and Mendgen (1984); Freytag and Mendgen (1991) reported that the wall of infection hypha is quite thin and covered with a matrix. Ehrahim *et al.*, (1985) and Harder *et al.*, (1986) reported that the inner wall layer of infection hyphae *of P. packyrhizi*, *U. viciae-fabae*, and *P. graminis* has high affinity to WGA.

Welter *et al.*, (1988) found tubular vesicular complex in intercellular hyphae and in haustoria. Health (1988) reported that infection starting from basidiospores of *Uromyces vignae* induces hypersensitive cell death as soon as they penetrate epidermal cells of an incompatible host plant. Boller and Metraux (1988) reported that Mannoprotein may cover the chitin and make the wall resistance to further attacked by chitinase and beta-1, 3-glucanases occurring in leaf apoplast. Mims *et al.*, (1989) found inner layer of infection hyphae of *P. arachidis* was covered with additional easily dicernible coatings.

Grignon and Sentenao (1991) reported that the physiochemical properties of the cell wall degrading enzymes of *Uromyces viciae-fabae* may be of critical importance in avoiding extended tissue damage and thus for establishment of biotrophy since pH of the apoplast was in weakly acidic range pH (5 - 6.5). Frittrang *et al.*, (1992) reported that in the biotrophic fungus *U. viciae-fabae*, pectin esterase isoenzymes have been separated by chromatofocusing and these forms of enzymes, showing pH of 8.4, 5.7 and 4.7 were detected 9 hours after inoculation when young hypha was formed. Heiler *et al.*, (1993) reported that at the stages of infection hypha differentiation activity of neutral cellulases increased dramatically and this increase continued until haustorial mother cell formed.

Haustorial formation

Borland et al., (1980) studied the diameter of aecial haustorium at the site of host wall penetration was greater than telial haustorium, and aecial haustorium was hyphal in appearance while the telial haustorium posses a slender neck region with darkely staining neck band a lobbed haustorial body. The aecial haustorium characteristically possesses a septum near its proximal end while such a septum was absent from telial haustorium. Voegele et al., (2001) reported that fungal biotrops differentiate specialized infection structures within the living cells of their host there haustoria have been linked to nutrient uptake ever since their discovery. uptake sugars from the host (Vicia fabae) to the rust fungus (Uromyces fabae) seems to be occurred largely through haustorial complex. Quilliam and shattock (2003) reported that Uromvces ficariae (microcyclic) and U. dactylidis (macrocyclic, heteroecious) both produced vermiform and largely indeterminate intracellular hyphae typical of M- haustoria associated with telial and aecial galls, respectively on their common host Ranunculus ficaria. Similar M-haustoria were observed in telia galls of Puccinia tunida (microcyclic) on Conopodium nrajus in host tissue affected by nine other microcyclic spp and in aecial galls of P. smyrni (demicyclic, autoecious) on Smyrinium alusatrum. To identify gene expressed during biotrophic growth, EST sequencing was performed with a haustorium cDNA library from Uromyces fabae.

Development of aecium cup

Orientation of aecium cup

Spiers and Hopcraft (1985) first indicated aecial development was the formation of rows of vertically orientated club-shaped cells within mesophyll tissue. These elongated and divided forming a single terminal peridial cell and an intercalary cell, which later broke down. Peridial cells thus formed a protective layer over underlying aeciosporophores. Developing columns of aeciospore initials later obscured all evidence of peridial cell formation. Peridial cells were dikaryotic and were readily distinguished from aeciospores by their solitary nature and dagger-like ornamentation. These aecia were subepidermal and developing aeciospores were covered with single layer of dikaryotic peridial cells. Aeciospores formation was meristem arthosporic Aeciospores were globose to subglobose and verrucose with cog like knobs superficially embedded in the secondary cell wall reported in Malampsora larici

Formation of peridium layer

Rijkenberg and Truter (1974) reported sporogenus cell or aeciosporophores of aecium eventually formed at the base of aecial primordium and appear to arise from large multinucleate cells. The sporogenus cells produced in this way each contain two nuclei that divide conjugately during formation of aeciospores initials two of the daughter nuclei remain in the sporogenus cell while the other two move into aeciospore initial. After the initial is delimited from the mother cell by a septum, the nuclei in the initial divide again and a transverse septum separates the initial into binucleate aeciospores and a small wedge-shaped, binucleate, sterile, intercalary or disjunctor cell (Rijkenberg and Truter, 1974). In most of species of rust fungi the peripheral cells of aecial base undergo successive divisions to produce a wall that surrounds the spore chains. Peterson (1974) reported that aecia of M. medusae and M. pinitorque were peridiate. Wilson and Henderson (1966) reported that there was no evidence for the peripheral paraphyses like hyphae neither united to form a rudimentary peridium nor were peridia formed as early products of basal cells which later formed

aeciospores as speculated for Melampsora species by (Peterson, 1974).

Formation of aeciospores chain

Hughes (1970) reported that aeciospores were meristem arthosporic. Rijkenberg and Truter (1974) reported that aeciospores are not annellophoric in P. sorghi and also reported that cell analogous to the multinucleate fusion cell was observed in the base of aecial storma.

However, in *M. epitea* and *M. larici-populina* the cell were less extensive than those of P. sorghi. Rijkenberg and Truter (1975), Littlefield and Health's (1979) reported that aeciospores ornamentation developed as outerlined. Holm and Tibbell (1974), Moore and McAlear's (1961) studied the vertucose aeciospores of *M. epitea* and *M. larici-populina* with their cog like knobs were similar to those of P. recondita. Gold and Littlefield (1979) reported that aeciospores of M. lini, in contrast, were coarsely verrucose and spine were wart-like.

References

1. Alexopolus, C. J., Mims, C. J. and Blackwell, M. (1996) Introductory Mycology. John Wiley and sons. Inc. 869 pp.

2. Arthur, J. C. and Cummins, G. B. (1962). Manual of rusts in and Canada. New York, Hafner Publishing United States Company.

3. Baruah, H. K. (1980). Text Book of Plant Pathology. Oxford and IBH. New delhi.

4. Bandre, A .and Pande, P. C. (2006). Introductory Botany. Rastogi Publication Meerut - Newdelhi.

5. Beckett, A., Tatnell, J. A. and Taylor, N. (1990). Adhesion and pre-invasion behavior of urediospores of Uromyces viciaefabae during germination on host and synthetic surface. Mycological Research 94: 865-875.

6. Bilgarmi, K. and Jamaluddin, S. and Rizvi, M. A. (1979). List of Fungi Part- I. List of References. Today and Tomarrow's Printers and Publishers New Delhi 110005 264 pp.

7. Borland, J. and Mims, C. W. (1980). An ultrastructural comparison of the aecial and telial haustoria of the autoecious rust Puccinia podophylli.Mycologia, Vol 72: 767-774.

8. Boller, T. and Metraux, J. P. (1991). Extracellular localization of chitinase in cucumber. Physiological and Molecular Plant Pathology 33: 11-16.

9. Bourett, T. M. and Howard, R. J. (1991). Ultrastructural

Immunolocalization of actin in fungus. Protoplasma 163: 199-202.

10. Butler, E. J. (1912). Fungi and Diseases in Plants. Thatcher, spink, co. Calcutta: 547 pp.

11. Chand, R., Srivastava, C. P., Singh, B. D. and Sarode, S. B. (2004). Identification and characterization of slow rusting components in pea (Pisvum sativum). Genetic Resources and Crop Evoluation 00: 1-6,

12. Chand, R., Srivastava, C. P. and Kushwaha, C. (2004). Screening technique for pea (Pisvum sativum) genotype against rust disease (Uromyces fabae). Indian Journal of Agricultural Sciences 74: 166-167.

13. Chaubal, R., Wilmot, V. A. and Wynn, W. K. (1991). Visualization adhesives and cytochemistry of the extracellular matrix produced by Urediospores germtube of Puccinia sorghi. Canadian Journal of Botany 69: 2044-2054.

14. Chauhan, M. P, Singh, I. S. and Singh, R. S. (1996). Genetics of Rust Resistance in lentils (Lens culinaris). Indian Phytopathology 49: 469-475.

15. Chen, C. Y. and Heath, M. C. (1992). Effect of stage of development of the cowpea rust fungus on the release of cultivar-specific elicitors of necrosis. Physiological and Molecular Plant Pathology. 40: 23-30.

16. Chong, J., Harder, D. E. and Rohringer, R. (1981). Ontogeny or mono and dikaryotic rust haustoria Cytochemical and ultrastructural studies. Phytopathology 71: 975-983.

17. Conner, R. L. and Bernier, C. C. (1982). Host range of Uromyces viciae-fabae. Phytopathology 72: 687-689.

18. Davies, M. E., Butler, G. M. (1986). Development of infection structure of the rust *Puccinia porri* on leek leaves. Transactions of British Mycological Society 86: 475-515.

18. DeBary, A. (1862). Morphologie and Physiologie- der plize Flechten and Myxomyceter.

19. Deising, H., Jungblut, P. R., and Mendgen, K. (1991). Differentiation related proteins of the broad bean rust fungus *Uromyces viciae-fabae* as revealed by high resolution two dimensional polyacrylamide gel electrophoresis. Archives of Microbiology 155: 191-198.

20. Deising, H. and Mendgen, K. (1992). Developmental control of enzymes production and cellwall modification in rust fungi and defence reactions of host plant. In U,T, Stahl P, eds Molecular biology of filamentous fungi Weinheim, Newyork, Basel, Cambridge: VCH verlagagessells chaft mbH: 27-44.

21. Deising, H., Nicholoson, R. L., Haug, M., Howard, R. J. and Mendgen, K. (1992). Adhesion pad formation and involvement of cutinase and esterase in the attachment of urediospores to the host cuticle. The Plant cell 4: 1101-1111.

22. Deising, H., Nicholson, R. L, Haug, M., Howard, R. J. and Mendgen, K. (1992). Adhesion Pad formation and involvement of cutinase and esterase in the attachment of urediospores to the host cuticle. The Plant cell 4: 1101-1111.

23. Dyke, C. G. and Mims, C. W. (1991). Ultrastructure of conidia, conidium germination and appresorium development in the plant pathogenic fungus *Collectotricum trieneatum*. Canadian Journal of Botany 69: 2455-2467.

24. Emeran, A. A., Sillero, J. C., Niks, R. E and Rubiales, D. (2005). Infection structures of host-specialized isolates of *Uromyces viciae-fabae* and of other species of *Uromyces* infecting leguminous crops. Plant Disease 89: 17-22.

25. Freytag, S., Bruscaglioni, I., Gold, R. E., and Mendgen, K. (1988). Basidiospores of rust fungi (*Uromyces species*) differentiate infection structures invitro. Experimental Mycology 12: 275-283.

26. Freytag, S. and Mendgen, K. (1991). Surface carbohydrate and cellwall structure of invitro induced urediospores infection structure of *Uromyces viciae-fabae* before and after treatment with enzymes and alkali. Protoplasma 161:94-103.

27. Freytag, S., Bruscaglioni, L., Gold, R. E. and Mendgen, K. (1988). Basidiospores of rust fungi (*Uromyces species*) differentiate infection structures in vitro. Experimental Mycology 12: 275-283.

28. Frittrang, A. K., Deising, H. and Mendgen, K. (1992). Characterization and partial purification of peptinesterase, differentiation specific enzymes of *Uromyces viciae-fabae*. Journal of General Microbiology 138:2213-2218.

29. Gaumann, E. A. (1998). Comparative Morphology of Fungi. Translated by Caroll William Dodge, Biotech Books, Delhi pp: 563.

30. Gold, R. E. and Littlefield, L. D. (1979). Light and scanning electron microscopy of the telial, pycnial and aecial stages of Melampsora lini. Canadian Journal of Botany 57:629-638.

31. Gold, R. E. and Mendgen, K. (1984). Cytology of basidiospore germination, penetration, early colonization of

Phaseolus vulgaris by Uromyces appendiculatus var. appendiculatus. Canadian Journal of Botany 62: 1989-2002.

32. Grambow, J. J. (1977). The influence of violatile leaf constituents on the invitro differentiation and growth of *Puccinia graminis f.sp.* tritici. Zeitschrift fur Pflanzenphysiologie 85: 361-372.

33. Grignon, C. and Sentenao, H. (1991). pH and ionic condition on apoplast. Annual Review of Plant Physiology and Plant Molecular Biology 42: 103-128.

34. Hamer, J. E., Howard, R. J., Chumley, F. G., and Valent, B. (1998). A mechanism for surface attachment in spores of a plant pathogenic fungus. Science 239: 288-290.

35. Harder, D. E., Chong, J., Rohringer, R. and Kim, W. K. (1986). Structure and cytochemistry of walls of urediospores germtubes and appressoria of Puccinia graminis tritici. Candian Journal of Botany 64: 476-458.

36. Harder, D. E. and Chang, J. (1991). Rust haustoria. In: Mendgen K, Lescmann D-E, eds. Electron microscopy of plant pathogens. Berlin: Springer Verlag, 235-250.

37. Heath, I.B. and Health, M. C. (1976). Ultrastructure of mitosis in the cowpea rust fungus *Uromyces phaseoli var. vignae*. The Journal of Cell Biology 70: 592-607.

38. Heath, M. C. (1989). In vitro formation of haustoria of the cowpea rust fungus *Uromyces vignae* in the absence of a living plant cell. Light microscopy. Physiological and Molecular Plant Pathology 35: 357-366.

39. Heath, M. C. (1990). In vitro formation of haustoria of the cowpea rust fungus *Uromyces vignae* in the absence of a living plant cell 2nd Electron microscopy. Canadian Journal of Botany 68: 357-366.

40. Heath, M. C. and Heath, I. B. (1975). Ultrastructural changes associated with the haustorial mother cell septum during haustorial formation in *Uromyces phaseoli var. vigane*. Protoplama 84: 279-314.

41. Heath, M. C. and Perumalla, C. J. (1988). Haustorial mother cell development of *Uromyces vignae* on collodium membrane. Canadian Journal of Botany 66: 736-741.

42. Heiler, S., Mendgen, K. and Deising, H. (1993). Cellulolytic enzymes of the obligatory biotrophic rust fungus *Uromyces viciae-fabae* are regulated differentiation-specifically. Mycological Research 97: 77-85.

43. Hiratsuka, Y., Morf, W. and Powell, J. M. (1966). Cytology of the aeciospores and aeciospores germtubes of *Peridermium harknessii* and *P. stalactiforme* of *Cronartium coleosporioides* complex. Canadian Journal of Botany 44: 1639-1643.

44. Hickman, M. J. and Epstein, L. (1988). *Necteria haematococca* macroconidia attach to plant surfaces. Phytopathology 78: 1523.

45. Hoch, H. C. and Staples, R. C. (1983). Ultrastructural organization of the non-differentiated urediospores germling of *Uromyces phaseoli*. Mycologia 6: 1209-1213.

46. Hoch, H. C. and Staples, R. C. (1987). Structural and chemical changes among the rust fungi during development. Annual Reviews of Phytopathology 25: 231-247.

47. Hoch, H. C. and Staples, R. C. (1984). Evidence that cyclic AMP initiates nuclear division and infection structure formation in the bean rust fungus, *Uromyces phaseoli*. Experimental Physiology 8: 37-46.

48. Holm, L., Tibell, L. (1974). Studies of the fine structure of aeciospores III. Aeciospore ontogeny in *Puccinia graminis*. Svensk Botanisk Tidskrift 68: 136-152.

49. Hughes, S. J. (1970). Ontogeny of spore forms in Uredinales. Canadian Journal of Botany 48: 2147-2157. 50. Joseph, M. E. and Hering, T. F. (1996). Effect of environment on spores germination and infection by broad bean rust (*Uromyces viciae-fabae*). Journal of Agricultural Science (Cambridge) 128:73-78.

51. Kaminskyj, S. W. G. and Day, A. W. (1984). Chemical induction of infection structures in rust fungi 2nd Inorganic ions. Experimental Mycology 8: 193-201.

52. Kapooria, R .G. and Mendgen, K. (1985). Infection structures and their surface changes during differentiation in *Uromyces fabae*. Phytopathologishe Zeitschrift 113: 327-323.

53. Kapooria, R. G. and Sinha, S. (1966). Studies on host range of *Uromyces fabae* (Pers) deBary. Indian Phytopathology 19: 224-230.

54. Kapooria, R. G. and Sinha, S. (1971). Further studies on host spectrum of *Uromyces fabae* (Pers) debary in India. Indian Phytopathology 24: 170-171

55. Kemen, E., Ariane, C., Rafiqi, M., Hempel, U., Mendgen, K., Hahn, M. and Voegele, R. T. (2005). Identification of protein from Rust Fungi transferred from haustoria into infected Plant cells. Phytopathology 153:43-47.

56. Kispatic, J. (1949). Prilog poznovanju biologiei suzbijanaja bobve rdje *Uromyces fabae* (Pers) debary *f.sp.* viciae-fabae debary. Annuals of Transaction Agriculture Society

Koch, E., Ebrahim, N. F. and Hoppe, H. H. (1983). Light and electron microscopic studies on the development of soybean rust (Phakopsora pachyrhizi. Syd) in susceptible soybean leaves. Phytopathologische Zeituchrift 106: 302-320.

57. Kushwaha, C., Chand, R., and Srivastava, C. P. (2006). Role of aeciospores in outbreak of pea (Pisvum sativum) rust (*Uromyces fabae*). European Journal of Plant Pathology 115: 323-330.

58. Kushwaha, C., Srivastava, C. P., Chand, R. and Singh, B. D. (2007). Identification and evaluation critical time for assessment of slow rusting in pea against *Uromyces fabae*. Field Crop Research 103: 1-4.

59. Kushwaha, C. (2006). Characterization and inheritance of slow rusting components in pea. Ph.D Thesis. B. H. U., Varanasi Kwon, E., Hoch H. C. and Aist, J. R. (1991). Initiation of appresorium formation in *Uromyces appendiculatus* organization of the apex and the responses involving microtubules and apical vesicles. Canadian Journal of Botany 69: 2560-2573.

60. Littlefield, L. J., Heath, M. C. (1979). Ultrastructure of rust fungi. London, Academic Press.

61. Macko, V., Renwick, J. A. A. and Rissler, J. F. (1978). Acrolein induces differentiation of infection structures in the wheat stem rust fungus. Science 199: 442-443.

62. Maheswari, R., Hildebrandi, A. C. and Allen, P. J. (1967). 63. The cytology of infection structure development in uredospore germ tubes of *Uromyces fabae var. typica* (Pers) Wint. Canadian Journal of Botany 45: 447-450.

64. Mendgen, K. (1973). Microbodies (glyoxysomes) in infection structure of *Uromyces phaseoli*. Protoplasma 78: 477-482.

65. Meinecke, E. P. (1929). Experiments with repeating pine rusts. Phytopathology 19: 327-342.

66. Mims, C. W., Taylor, J. and Richardson, E. A. (1989).Ultrastructure of the early stages of infection of peanut leaves by the rust fungus Puccinia arachidis. Canadian Journal of Botany 50: 767-786.

67. Mitter, J. H. and Tandon, R. N. (1930). Fungi flora of Allahabad India. Journal of Indian Botanical society 9: 190-196.

Moore, R. T. and McAlear, J. H. (1961). Fine structure of mycota 8. On the aecidial stage of Uromyces caladii. Phytopathologische zeitschrift 42 : 297-304.

68. Nicholson, R. L. and Epstein, L. (1991). Adhesion of fungi to the plant surface: Prerequisite for Pathogenesis In: cole G T, Hoch H C eds. The fungal spores and disease initiation in plants and animals. New York: Pienum press 3:23.

69. Nighswander, J. E. (1963). Studies in forest tree rusts in Alberta, p. 118. In Canada Department of Forestry, Forest Entomology and Plant Pathology Branch, Annual Report, Year ended March 31, 1963.

70. Peterson, R. S. (1974). Rust fungi with caeoma-like sori on conifers. Mycologia 66: 242-255.

71. Persoon, D.C. H. (1801). Synopsis methodica fungorum 1: 224.

72. Prasada, R. and Verma, U. N. (1948). Studies on lentil rust, *Uromyces fabae* (Pers) deBary in India. Indian Phytopathology 1: 142-146.

73. Patel, M. K. (1934). Indian Bulletin and Plant Protection (8): M199-200.

74. Pavgi, M. S. and Upadhyay H. P. (1966). Parasitic Fungi from North India IV Mycopathology et. Mycological Applications 30: 257-260.

75. Quilliam, R. S. and Shattock, R. C. (2003). Haustoria of microcyclic rust fungi Uromyces ficariae and Puccinia tumida and other gall forming species Uromyces dactylidis (macrocyclic) and *Puccinia smyrnii* (demicyclic). Plant Pathology 52:104-113.

76. Read, N. D., Kellock, L. J., Knight H. and Trewaves A. J. (1992). Contact sensing during infection by fungal pathogen. In: Callow J. A., Green J. R., eds. Perspectives in plant cell recognition. Cambridge University Press, 137-172.

77. Rijkenberg, F. H. J. and Tructer, S. J. (1973). Sporogenesis of *Puccinia sorghi* on alternate host. Proceedings of the Electron Microscopy Society of South Africa 3: 3-4.

78. Rijkenberg, F. H. J. and Tructer, S. J. (1974). The ultrastructure of sporogenesis in the pycnial state of *Puccinia sorghi*. Mycologica 66:319-326.

79. Rijkenberg, F. H. J. and Tructer, S. J. (1974). The ultrastructure of the *Puccinia sorghi* aecial stage. Protoplasma 81: 231-245.

80. Rijkenberg, F. H. J. and Tructer, S. J. (1975). Cell fusion in the aecium of *Puccinia sorghi*. Protoplasma 83: 233-246.

Roy, T. C. (1949). Fungi of Bengal: Directorate of Agriculture. Govt. of West Bengal.

81. Rubiales, D. and Sillero, J. C. (2002). *Uromyces viciae-fabae* haustorium formation in susceptible and resistance faba bean lines. European Journal of Plant Pathology 109: 71-73.

82. Staples, R. C. (2000). Research on rust fungi during the twentieth century. Annual Review of Phytopathology 38:49-69.

83. Singh, S. J. and Sokhi, S. S. (1948). Evaluation of pea cultivar to *Uromyces viciae-fabae*. Indian Phytopathology 21: 85.

84. Singh, S. J. and Sokhi, S. S. (1980). Pathogenic variability in *Uromyces fabae*. Plant Diseases 64: 671-672.

85. Singh, R. S. (1999). Plant Diseases Oxford IBH Pub. NewDelhi pp: 686.Singh, C., Singh P., and Singh, R. (2003). Modern Techniques of Raising Field Crops IInd Edition. Oxford IBH Pub. NewDelhi pp 219-221.

86. Sillero, J. C., Moreno, M. T. and Rubiales, D. (2000). Characterization of new sources of resistance to *Uromyces viciae-fabae* in a germplasm collection of *Viciae faba*. Plant Pathology 37: 123-125.

87. Sokhi, S. S. (1984). A distinct Physiologic race of *Uromyces viciae-fabae* on Sweet pea. Indian Phytopathology 49: 387-388.

Spiers, A. G. and Hopcraft, D. H. (1985). Ultrastructural studies of the spermatial and aecial stages of *Melampsora laricipopulina* and *Melampsora* epitea on Larix *deciduas*. New Zealand Journal of Botany 23: 101-116.

88. Swann, E. C. and Mims, C. W. (1991). Ultrastructure of freeze-substituted appressoria produced by aeciospore gremlings of the rust fungus Arthuriomyces peckianus. Canadian Journal of Botany 69:1655-1665.

89. Ture, R. P. (1938). Gall development on Pinus sylvestris attacked by wood-gate Peridermium, and morphology of the parasite. Phytopathology 58: 309-315.

90. Uppal, B. M. (1933). International Bulletin of crop protection M 7:M103 and M746.

91. Vijaylaxami, S., Yadav, K., Kushwaha, C., Sarode, S. B., Srivastava, C. P., Chand, R. and Singh, B. D. (2005). Identification of RADP marker linked to the rust (*Uromyces fabae*) resistance gene in pea (*Pisvum sativum*). Euphytica 144: 265-274.

92. Voegele, R. T., Hahn, M., Lohaus, G., Link, T., Heiser, I., and Mendgen, K. (2005). Possible roles for Mannitol and

Mannitol Dehydrogenase in the Biotrophic plant pathogenic *Uromyces fabae*. Plant Physiology 137:190-198.

93. Voegele, R. T., struck, C., Hahn, M. and Mendgen, K. (2001). The role of haustoria in sugar supply during infection of broad bean by rust fungus *Uromyces fabae*. PNAS 98: 8133-8138.

94. Welter, K., Muller, M. and Mendgen, K. (1988). The hyphae of Uromyces appendiculatus within the leaf tissue after high pressure freezing and freeze substitution. Protoplasma 147:91-99.

95. Wilson, M. and Henderson, D. M. (1966). British rust fungi. London, Cambridge University Press.

96. Xue, A. G. and Warkentin, T. D. (2002). Reactions of field pea varieties to three isolates of *Uromyces fabae*. Canadian Journal of Plant Sciences 82: 253-255.

97. Xu, H. and Mendgen, K. (1991). Early events in living epidermal cells of cowpea and broad bean during infection with basidiospores of the cowpea rust fungus. Canadian Journal of Botany 69: 2279-2285.

98. Zhou, X. L., Stumpf, M. A. and Hoch, H. C. (1991). A mechanosensitive channel in whole cells and in membrane patches of the *fungus Uromyces*. Science 235:1415-1417.