Available online at www.elixirpublishers.com (Elixir International Journal)

Pharmacy

Elixir Pharmacy 41 (2011) 5657-5663

Optimizing antimicrobial drug use in surgery: an intervention strategy in a Sudanese hospital to combat the emergence of bacterial resistant

Salah Ibrahim Abd Alrhman Kheder

National College of Medical & Techenical Sciencies- Pharmacy program, Khartoum - Sudan, 3783 Khartoum.

ARTICLE INFO

Article history: Received: 29 August 2011; Received in revised form: 16 November 2011; Accepted: 27 November 2011;

Keywords

Antibiotic resistance, Antimicrobial cycling, Antimicrobial rotation, Surgical prophylaxis.

ABSTRACT

Antimicrobial control programs are widely used to decrease antibiotic utilization, but effects on antimicrobial resistance and outcomes for patients remain controversial. The purpose of this study was to determine the impact of rotation of antibiotic classes used as empirical surgical prophylaxis on the emergence of bacterial resistance organisms and antibiotics drug use when compared with non-rotation period. Three core, broad spectrum agents (Cephalosporins, beta-lactam-inhibitors, and fluoroquinolones) were selected for inclusion in the quaternary rotation for 21 months, based on prior 8 months baseline data from GIT and urology surgical wards in Ibn Sina hospital. Intensive surveillance done for patients admitted to the selected settings. 1681 surveillance samples obtained from 2359 eligible inpatients admitted to hospital from Jan 2008 to May 2010. A significant reduction in the percentage of positive growth had been observed with antibiotic rotation for both wards from 65% and 49% in baseline to 59% and 33% in rotation (1) and 25% and 33% in rotation (2) in GIT and urology ward respectively ($p \le 0.0001$). As general there was a divergent effect of the antimicrobial rotation on the prevalence of resistance among G+ve and G-ve bacteria. We concluded that antimicrobial drug use in surgical departments could be optimized after implementation of antimicrobial cycling policy, and associated in reduction in the incidence of infectious mortality and morbidity but stabilize antibiotic resistance, without significant reduction.

© 2011 Elixir All rights reserved.

Introduction

Surgical procedures, by their very nature, interfere with the normal protective skin barrier and expose the patient to microorganisms from both endogenous and exogenous sources. Infection resulting from this exposure may not be limited to the surgical site but may produce wide spread systemic effects. Traditional control measures include sterilization of surgical equipments, disinfection of the hand and skin, use of antibiotics.1 prophylactic The prophylactic antibiotic administration is a complementary to surgical treatment of site infections, contributing substantially to minimizing of complications, morbidity, and death.² The basic principle of antibiotic prophylaxis in surgery is to achieve adequate serum and tissue drug levels that exceed, for the duration of the operation, the MICs for the organisms that are likely to be encountered during the operation.³ The selection of appropriate antimicrobial agents depends on the identification of the most likely pathogens that are associated with a specific surgical operation. The prolonged use of prophylactic antimicrobials is associated with emergence of resistance bacterial organisms.⁴

Resistant organisms pose a grave threat to hospitalized patients as their prevalence increases and antibiotic options narrow, mandating aggressive strategies to control their elaboration and spread. As antibiotic usage has been implicated as a key factor in the development of resistance⁵, various techniques of formulary restriction⁶, decision support tools⁷, antibiotic therapy⁹ and antibiotic cycling or rotation¹⁰ have been advocated as means to control potentially unnecessary and inappropriate antibiotic usage. There has been increasing interest

in antibiotic cycling or rotation, as clinicians seek novel methods to combat the epidemic emergence of resistant organisms in hospitals around the world.¹¹ The efficacy of cycling or rotating antibiotic classes in reversing or forestalling antimicrobial resistance remains controversial. Whereas some studies have implied improvements in antimicrobial resistance patterns or outcomes with cycling, other trials have been largely negative or have even led to worsening of resistance¹².

Through the use of a predetermined quarterly schedule of empiric antibiotics optimally as a prophylactic pre and postoperatively, we hypothesized that rotation could be associated with significant decreases in rates of infection, resistant gram-negative and gram-positive organisms and antibiotics consumption when compared with non-rotation period.

Method:

Study population:

This prospective study was performed in two surgical wards (Gastrointestinal-GIT- surgical ward and Urology surgical ward) in Ibn Sina hospital, 132-beds secondary teaching hospital- Khartoum state capital of Sudan, from Jan 2008 until May 2010. The population of the study was sequential. Patients admitted to GIT and Urology surgical wards in Ibn Sina hospital for \geq 48 hours were eligible for the study, and followed prospectively until discharge or death. The included inpatients were patients that underwent into surgical operation.

Study protocol implementation:

This was a prospective before-and-after study. A detailed account implementation has been previously described.¹³ Briefly, antibiotic rotation protocol was implemented in



September 1, 2008, as a local hospital policy for antibiotic prophylaxis pre and postoperatively. Baseline data were collected for 8 months (Jan 1 to August 30, 2008). During the baseline period, the prescription of antibiotics for surgical prophylaxis for the antibiotic coverage was at the discretion of the ordering surgeon. After the baseline observation period, an antibiotic-cycling protocol was implemented. Three antibiotics, Cephalosporins (CEF), Co-amoxiclave (AMC) and Ciprofloxacin (CIP) were empirically cycled as primary antibiotics for surgical prophylaxis every 3 to 4 months over a 2year period. These three cycled drugs were systemically rotated twice, with the cycled drug changing every 4 months in the first year (rotation 1) and 3 months in the second year (rotation 2). The goal of this rotation was to direct quarterly antibiotic class heterogeneity in an effort to avoid resistance -selective pressure. Data collection and analysis:

Antibiotic susceptibility data for gram positive and gram negative bacteria were collected 8 months before (baseline period), and 21 months after (intervention period), September 1, 2008. Specimen for culture and sensitivity were collected twice times per week from each ward (As surgical operations done twice/week for each ward), from eligible patients as surgical swabs from GIT & urology wards or urine samples from urology ward only and sent to the hospital laboratory for culture and sensitivity tests. Also during this period demographic, clinical and pharmacological data were obtained. The following aspects of antimicrobial prophylaxis were audited: antibiotic choice, duration, dose, interval between doses. Wound class, physical condition of the patient according to classification of the American Society of Anesthesiologist (ASA) was recorded.¹ Adherence to local guidelines for antimicrobial prophylaxis was reviewed for intervention period. Data were collected by infection control practitioner from medical and nursing records, and medication chart, using standardized form. Data collection was validated and entered in WHONET¹⁴ database at monthly base by the investigator. The data collected were analyzed using WHONET analysis software, Excel 2007 and SPSS version 16.0. Antimicrobial drug consumption received by the patients prophylactically was converted into Defined Daily Dose (DDD). Quantitative use was calculated and compared as (DDD/100-bed days)¹⁵. Prior to initiation of the study, ethical approval was obtained from Medical Ethical Committee Ministry of Health and also hospital approval was obtained. Considering the observational nature of the study, the use of conventional antibiotic therapy, so there is no need to obtain informed consent from the patients.

Results:

A total of 2359 patients were eligible to be included into the study according to the study criteria. 2329 (98.7%) of them were underwent into surgical operations, 637 (27%) GIT and 1692 (73%) urological operations. About 68% (1583) of them were male and the mean age range between $47 \pm 15.2 - 53.4 \pm 17.02$ for GIT ward 41±22.6 - 44±21.6 for urology ward. Different reasons for surgical operations and underline diseases for admission to both surgical wards, but the main reasons were stones and cancer of GIT and urology system. The length of stay decreased from pre-intervention period to post-intervention period for each ward, but was not statistically significant reduction (GIT 13.3± 11.8 Vs 9.6± 8.7 $p \le 0.229$; Urology 11.9 ± 12.42 Vs $7.1 \pm 5.5 \ p \le 0.204$). A decrease in the mortality rate was observed when comparing between the two study periods for each ward, but also without significant difference. The detailed and other characteristics of the study populations before and after intervention study periods were shown in Table (1):

Total antibiotics used during the study period in GIT ward was 81.4 DDD/100 bed-days and in urology ward was 193.05 DDD/ bed-days. Total protocolized antibiotics used were 47.5 DDD/100 bed-days in GIT ward and 168.6 DDD/100 bed-days in urology ward. However, mean percentage of patients received the protoclized antibiotic decreased in rotation (2) compared to rotation (1) by 20% in GIT and 17% in urology surgical wards. The median duration of antibiotic treatment days increased from 3 days to 4 days in GIT ward, while decreased in urology ward from 3 days to 2 days. 1681 surveillance samples obtained from 2359 eligible inpatients admitted to the Ibn Sina hospital throughout study period from Jan 2008 to May 2010. Of these samples 345 (20.5%) obtained from GIT ward as surgical and wound swabs, 1336 (79.5%) samples obtained from urology surgical ward (1197 urine samples and 139 surgical swabs). Specimen obtained from patients during the post-intervention periods was more than pre-intervention period, but a significant reduction in the percentage of positive growth had been observed with antibiotic rotation for both wards from 65% and 49% in baseline to 59% and 33% in rotation (1) and 25% and 33% in rotation (2) in GIT and urology ward respectively ($p \le$ 0.0001). А substantial variation in incidence of colonization/infection rate was observed between the two surgical wards, while it decreased in GIT ward it increased in urology ward when compared between pre and after intervention periods.

Details of cycled antibiotics consumed in DDD during the non rotation and rotational periods in GIT and urology surgical wards were shown in figures (1):



Figure (1): Amounts of cycled antibiotics consumed in DDD in (a) GIT ward (b) urology ward per study periods

The most frequently categories of antibiotics prescribed throughout the study period were cephalosporins for both GIT and urology wards, and the use of cephlosporins were not completely restricted during any period throughout the study period in both GIT and urology wards, while amoxiclave and quinolones were completely restricted in some cyclic periods in GIT ward (data were not shown). Cefuroxime was the main cephalosporin antibiotic prescribed in GIT surgical ward constitute (47.24%), followed by ceftazidime (27.42%) and ceftrixone (25.34%), while in urology surgical ward the heaviest cephalosporin prescribed was ceftrixone (54.16%), followed by cefuroxime (29.37%) and the lowest was ceftazidime (16.47%). In 58% and 68% of all cases in GIT and urology wards respectively, antibiotics were compliant and prescribed according to the protocol, higher compliance rate was observed in first rotation compared to second rotation in both wards, with a significant difference (70% vs. 41.5% in GIT ward; P =0.0001, and 75% vs. 60.5% in urology ward; P = 0.0001). In every cycle (where it is not the on-cycled antibiotics), cephalosporins were the most frequent off-cycle drug to be prescribed in both GIT and urology ward.



Antibiotic resistance in (A) gram- positive (G+ve) and (B) gram-negative (G-ve) isolates per each rotational cycle in GIT surgical wards. AMC –Co-amoxiclave, CAZ = Ceftazidime, CRO = Ceftrixone, CXA = Cefuroxime, CIP = Ciprofloxacin, Linear = Linear Trendlines of resistance

Figures (2) and (3) illustrate the diversity of resistant G +ve and G-ve isolated by rotation cycles in GIT and urology surgical wards respectively. Pattern of drug resistance were observed to differ pre and after intervention. In figure 2 (A) there was a trend towards decreasing of G +ve resistance to the three major antibiotic used in GIT ward (indicated as linear trendlines), whereas in figure 2 (B) G -ve resistance increased towards Coamoxiclave and cephalosporins (represented by cefuroxime), with more dramatic decreased resistance towards ciprofloxacin in the same surgical ward. In urology ward the trend suggest that increase in resistance associated with antibiotic used except towards cephalosporins (represented by ceftrixone) where there was a slight decrease in both G+ve and G-ve bacteria as shown in figure 3 (A & B):



Figure (3) Antibiotic resistance in (A) gram- positive (G+ve) and (B) gram-negative (G-ve) isolates per each rotational cycle in urology surgical wards. AMC –Co-amoxiclave, CAZ = Ceftazidime, CRO = Ceftrixone, CXA = Cefuroxime, CIP = Ciprofloxacin, Linear = Linear Trendlines of resistance

Details of gram positive and gram negative antimicrobial susceptibilities before and after intervention periods had been shown in tables (2 and 3). In rotation 1, only one isolates exhibit significant reduction in resistance towards Co-amoxiclave in *Staphylococcus aureus* during Co-moxiclave (from 100% to 61.1%) in GIT surgical ward (table 2). While many isolates exhibit significant reduction in resistance during rotation 2 in both GIT and urology surgical wards (table 3):

Discussion:

From initial review we concluded that antimicrobial drug use in surgical departments could be optimized after implementation of antimicrobial cycling policy. The policy replaced a variety of antimicrobial use regimens, previously chosen on the basis of personal preferences and possibly the result of promotional efforts by pharmaceutical companies. The intervention succeeded in decreasing the mean percentage of patients received antibiotic prophylactically. Other indicators of satisfactory outcomes with the new policy were a decrease, length of stay and mortality. The number of isolates that were isolated from study surgical patients increased in rotational periods compared to baseline periods, this due mainly to the longer rotational period and active surveillance system during those periods. But a significant reduction in the percentage of positive growth, we also found a trend favoring a lower incidence of colonization/ infection rate with antibiotic rotation. Other interventional studies have similarly demonstrated decreases in infection rates without significant changes in patient mortality and length of stay.¹⁶⁻¹⁸

We have previously shown that there were pronounced reductions in overall antibiotic use and total protocolized antibiotic utilization represented as a reduction in DDD/100 beddays measurement.¹⁹ This considerable reduction, may actually overshadow any impact of cycling program may have had on the measured outcomes. Cephalosporins were the most often cycled antibiotics prescribed in both surgical wards during the whole study period. Cephalosporins are frequently used either alone or in combination with metronidazole as surgical prophylaxis. The over use of broad spectrum cephalosporins particularly ceftizidime, cefuroxime and ceftrixone have been implicated in the emergence of multidrug-resistant gram positive and gram negative bacteria.²⁰

Despite our study got some success to lower the amount of utilized antibiotics used, but it seems this reduction is not enough to decrease antibiotic resistance (i.e. still above the threshold point to reduce resistance). Trends of bacterial resistance as a group and by organisms to cycling antibiotics showed no much significant differences between the 2 years. Although resistant development against β-lactam and flourquinolone antibiotics is based on different mechanism, homogenous exposure to one of these classes did not prevent resistance development to other classes.²¹ Antibiotic cycling has been suggested as a method for decreasing or controlling resistance in microorganisms. In theory, the antibiotic agents undergo rotation in a given time period, altering resistance pressure in microbial environment. Bacteria with resistance to an agent would lose their growth advantage when the agent withdrawn from use, and exposure to other class of antibiotics would eliminate these resistant organisms. The present study shows some differences between theoretical considerations and daily clinical practice. However, the theoretical benefits of antibiotic cycling hold true in daily practice can only be effective by controlling confounding variables. Part of the difficulties in controlling confounding variables arises from the lack of randomization in such quasi-experimental studies (preintervention and post intervention). On the other hand high cross-resistance between cyclic antibiotics and multi-resistance strains carried out by patients admitted to both surgical wards overwhelmingly dominant in the study wards, this indicated by the persistent multi-resistance profile and absence of significant decrease in antibiotic resistance among most of the cyclic periods for gram positive and gram negative species. The problem of multi-drug resistance may well decrease the potential benefits of antibiotic cycling. Also surgeons' adherence to only the use of the cycled antimicrobial was poor and also erratic and this may have a big role in altering the result of our study. Numerous studies have examined different strategies of rotating an assortment of antibiotic classes, ultimately yielding divergent results.16, 22-

Certain limitations exist in our study design. In the first three cycles (rotation 1), cycles were 4 months in the length. During rotation 2, cycles were 3 months in length. While this change might shed light on the question of appropriate cycle

Salah Ibrahim/ Elixir Pharmacy 41 (2011) 5657-5663

length for a successful cycling protocol, it is also limits the generalizability of the data, but this mainly due to the research funding limitations and low adherence and cooperation from prescribe at rotation two mainly. Also we did not link and assessed the infection according to the clinical picture and depend only on colonization and pathogenic isolate cultures and this may over estimate infection rate.

Conclusion

Antibiotic policy and guidelines were important to optimize antibiotic drug use for surgical prophylaxis. The adherence to such guidelines must be improved, to achieve optimal adherence, antibiotic policy makers should develop evidencebased guidelines in collaboration with surgeons.

Acknowledgment

We would like to express our deep gratitude and thankful to Ibn Sina hospital administration and healthcare worker staffs in different hospital units (Pharmacy, infection control, microbiology lab, statistic department, GIT and urology wards) for enduring the different study- protocols enforced on them and maintaining high level of co-operation during the different study periods.

References

1. Vivian G. Loo. Infection control in surgical practice. ACS surgery: Principles and Practice, 2008; 5 (8):1-11.

2. Apostolos G. Lambaroudis, Savvas Papadpoulos, Michelle Chiristodulidou, Thomas Gerasimidis. Perioperative use of antibiotics in intra-abdominal surgical infections. Surgical infections, 2010; 11 (6): 535-44.

3. Bratzler D, Houck P. Antimicrobial prophylaxis for surgery: an advisory statement from the national surgical infection project. Clin Infect Dis,2004; 38: 1706-15.

4. Gyssens IC. Preventing postoperative infections: current treatment recommendations. Drugs 1999; 57:175-85.

5. Ausin DJ, Krisitinsson KG, Anderson RM. The relationship between the volume of antimicrobial consumption in human communities and the frequency of resistance. Pro Natl Acad Sci USA, 1999; 96:1152-56.

6. Quale J, Landman D, Saurina G,et al. Manipulation of a hospital formulary to control an outbreak of vancomycin-resistant enterococci. Clin infect Dis 1996; 23: 1020-25.

7. Evans RS, Petotnik SL, Classen DC, et al. A computerassisted management program for antibiotics and other antiinfective agents. N Engl Med 1998; 338: 232-238.

8. Fridikin SK, Steward CD, Edwards JR, et al. Surveillance of antimicrobial use and antimicrobial resistance in United States hospitals: Project ICARE phase 2. Project intensive Care Antimicrobial Resistance Epidemiology (ICARE) hospitals. Clin Infect Dis 1999; 29: 245-52.

9. Singh N, Rogers P, Atwood CW, et al. Short-course empiric antibiotic therapy for patients with pulmonary infiltrates in the intensive care unit. A proposed solution for indiscriminate antibiotic prescription. Am J Respir Crit Care Med 2000; 162: 505-11.

10. McGwan JE Jr. Strategies for study of the role of cycling on antimicrobial use and resistance. Infect Control Hosp Epidemiol 2002; 21 (1 Suppl): S36-S43.

11. Niederman MS. Appropriate use of antimicrobial agents: Challenges and strategies for improvement. Crit Care Med 2003; 31: 608-16.

12. Brown Erwin M. & Nathwani Dilip. Antibiotic cycling or rotation: a systematic review of the evidence of efficacy. J. Antimicrob. Chemother., (2005b); 55:6-9.

13. Salah I. Kheder, Idris Eltayeb, Sania A I Shaddad, Esam Kheder. Impact of Antibiotic cycling Policy in Antimicrobial

Resistance in, Two Sudanese Surgical Wards Settings: Prospective Longitudinal Interventional Study. Journal of pharmaceutical and Biomedical Sciences 2011; 5(7) : 1-10

14. O'breien Thomas F, Stelling John M & Eskildsen Ma. Using internet discussion of antimicrobial susceptibility databases for continous quality immprovement of the testing and management of antimicrobial resistance. Clin Infect Dis, 2001; 33 (Suppl 3): S118-123

15. White R L. How do measurements of antibiotic consumption relate to antibiotic resistance. IN M I, Gould , & W J, Van Deer Meer (Eds.) Antibiotic policies theory and practice New Yourk, Springer. 2005; 75-105

16. Raymond D. P, S.J. Pelletier, T. D. Crabtree, T.G. Gleason, L. L. Hamm, T.L. Pruett & R.G. Sawyer. Impact of a rotating empiric antibiotic schedule on infection mortality in intensive care unit. Crit Care Med, 2001; 29, 1101-08.

17. Michael G Hughes, Heather L. Evans, Tae W. Chong, Robert L. Smith & Robert G Sawyer. Effect of an intensive care rotating empiric antibiotic schedule on the development of hospital-acquired infections on the non-intensive care unit ward. Crit Care Med, 2004; 32:53-60.

18. Salah I. Kheder, Idris Eltayeb, Sania A I Shaddad, Esam Kheder. Effect of antibiotic rotation protocol on the development of hospital acquired infections in hospital surgical units in Sudan. *Sudan Medical Monitor* 2010; 5 (4) 165-173.

19. Salah.I. Kheder . Antibiotic Utilization and Prescribers Adherence measurements in a Sudanese Hospital Settings after Introducing Antibiotic Policy . Sudan Medical Monitor 2011; 6 (1) 39-48.

20. Palmer S.M., Kang S.L., Cappelletty & D.M. And Rybak M.J. Bactericidal killing activities of cefepime, ceftazidime, cefotaxime and ceftriaxone against S.aureus and b-lactamase producing strains of Enterobacter aerogenes and Klebsiella pneumoniae in an invitro infection model. Antimicrobial agent and chemotherapy. 1995;39: 1764-1771.

21. Van Loon Harald J., Vriens Menno R., Fluit Ad C., Troelstra Annet, Van Der Werken Christiaan, Verhoef Jan & Bonten Marc J. M. Antibiotic Rotation and Development of Gram-Negative Antibiotic Resistance. *Am. J. Respir. Crit. Care Med.* 2005; 171: 480-487.

22. Domiguez Ea, Smith Tl, Reed E, Sanders Cc & Sanders We A pilot study of antibiotic cycling in a hematology oncology unit. *Infect Control Hosp Epidemiol*, 2000; 21: S4-S8.

23. Hughes M. G., Evans H. L., Chong T. W., Smith R. L., Raymond D. P., Pelletier S. J., Pruett T. L. & Sawyer R. G. Effect of an intensive care unit rotating empiric antibiotic schedule on the development of hospital-acquired infections on the non-intensive care unit ward. *Crit Care Med.*, 2004; 32:53-60.

24. Fridikin K, Scott, Routine cycling of antimucrobial agents as infection-control measure. Clin Infect Dis, 2003; 36: 1438-1444.

25. Hala Badawi, Manal Saad, Diab, & Manal Elsaid . Impact of Antibiotic Policy in a Tertiary Care Research Institute Hospital in Egypt: Three Years Experience. Onternational Journal of Infection Control, 2007; 3: 1-7.

5660

Table 1: Demographics and characteristics of patients in GIT and Urology surgical ward before and after intervention

periods													
Period	Before interve	ntion	After Intervention										
	Baseline perio	d	First rotation		Second rotation	1							
Surgical ward	GIT	Urology	GIT	Urology	GIT	Urology							
Demographic data					1								
Study length	8 months	8 months	12 months	12 months	9 months	9 months							
Study time	Jan 1, 2008	– August 30,	September 1, 20	08 – August 30,	September 1, 2009 – May								
	2008		2009		2010								
Total number of admitted patients	344	577	1154	1441	950	1153							
Number of eligible patients for the study	195	365	272	739	739 177								
Number of Patients enrolled in surgical operations (%)	188 (97%)	348 (96%)	272 (100%)	733 (99%)	177 (100%)	611 (100%)							
Patient / days	3246	4769	11757	15031	8981	9251							
Number of patient received antibiotics preoperatively (%)	185 (98.4%)	324 (88.8%)	227 (83.5%)	638 (87%)	163 (92%)	588 (96%)							
Number of patient received antibiotics Postoperatively (%)	178 (95%)	334 (98%)	247 (91%)	712 (97%)	171(97%)	598 (98%)							
Total antibiotic consumption in DDD	311.5	267.6	536.9	2258.1	338.4	1696.6							
DDD/100 bed-days	15.9	32.98	38	68.2	27.5	91.9							
Mean percentage of patients received the protoclized	NA	NA	62%	65%	42%	48%							
antibiotic													
America	n Society of Ane	sthesiologist (A	SA) classification										
ASA (1), N (%)	144 (76.6%)	283 (81.3%)	244 (89.7%)	663 (90.4%)	139 (78.5%)	591 (96.7%)							
ASA (2), N (%)	44 (23.4%)	64 (18.4%)	28 (10.3%)	70 (9.6%)	36 (20.4%)	20 (3.3%)							
ASA (3), N (%)	0 (0%)	1 (0.3)	0 (0.0%)	0 (0.0%)	2 (1.1)%	0 (0.0%)							
	Woun	d classification											
Clean wound, N (%)	9 (4.9%)	3 (0.8%)	2 (0.7%)	36 (4.9%)	16 (9.1%)	444 (72.6%)							
Clean contaminated wound N (%)	163 (86.7%)	285 (78.1%)	251 (92.3%)	625 (85.3%)	151 (85.3%)	166 (27.2%)							
Dirty wound N (%)	16 (8.5%)	60 (16.4%)	19 (7%)	72 (9.8%)	10 (5.7%)	1 (0.2%)							
Prevalence of Colonization/infection													
Number of specimen obtained from eligible patients (%)	49 (26%)	93 (26%)	189 (69%)	641 (87%)	107 (60%)	602 (99%)							
Number of positive growth from cultured specimens (%)	32 (65%)	46 (49%)	95 (50%)	212 (33%)	27 (25%)	196 (33%)							
Rate of prevalence of colonization/infection per 1000	9.6	9.7	8.1	14.1	3.0	21.2							
patient/days													
P^*			$P1 \leq 0.0001$	$P1 \leq 0.0001$	$P1 \leq 0.0001$	$P1 \le 0.0001$							
					$P2 \le 0.0002$	<i>P2</i> ≤0.7278							

 $P^* = P$ -value, z-test for proportions. P1 = P-value between baseline and rotation (1) P2 = P-value between rotation (1) and rotation (2) NA = Not Applicable

Salah Ibrahim/ Elixir Pharmacy 41 (2011) 5657-5663

Table (2): Doreontage of isolates resistance to	walad antibiation during the 8 months before	and 12 months after (rotation 1) noriads
Table (2). Tercentage of isolates resistance to	ycieu antibiolics uuring the o months before	and 12 months after (rotation 1) perious

	No	of	Cyclic period (Rotation 1)																			
	isolates		Cephalos	porins per	iod (CEP)				Co-am	oxiclave pe	riod (AM	(C)			Quinolones period (CIP)							
Organism (Give and Gve)			CEP(R%)		AMC (R%)		CIP(R%)		CEP	(R%)	AMO	C (R%)	CIP (R%)		CEP (R%)		AMC (R%)		CIP (%)			
Patient location			D (D.C		D (D.C		D.C		D (D.C		D (D C			
	D - f		Before		Before	e After	Before		Before		Before	•	Before		Before		Before		Before	After		
	After		Alter				Alter		Alter		Alter		Alter		Alter		Alter					
Staphylococcus aureus	711101																					
GIT ward	14	47	97.6%	87.3%	100%	62.5%	72%	90.9%	97.6%	70.7%	100%	61.1%*	72%	84.2%	97.6%	96.7%	100%	78.6%	72%	100%		
Urology ward	13	20		100%	100%	100%	92.3%	100%	100%	100%	100%	100%	92.3%	100%	100%	91.7%	100%	75%	92.3%	91.7%		
			100%																			
Staphylococcus saprophyticus																						
GIT ward	2	2	100%	100%	50%	100%	0%	100%	100%	NA	50%	NA	0%	NA	100%	NA	50%	NA	0%	NA		
Urology ward	2	1	100%	100%	100%	0%	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA		
Staphylococcus epidermidis																						
GIT ward	1	2	100%	NA	NA	NA	0%	NA	100%	NA	NA	NA	0%	NA	100%	50%	NA	100%	0%	50%		
Urology ward	3	0	77.8%	NA	33.3%	NA	66.7%	NA	77.8%	NA	33.3%	NA	66.7%	NA	77.8%	NA	33.3%	NA	66.7	NA		
																			%			
Streptococcus sp.																						
GIT ward	0	2	NA	100%	NA	100%	NA	100%	NA	NA	NA	NA	NA	NA	NA	100%	NA	NA	NA	100%		
Urology ward	0	2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	100%	NA	NA	NA	100%		
Entercoccus faecalis	0	0	NT A	NT 4	NT 4	NT A	NT 4	NT A	NT 4	NT 4	NT A	NT A	NT A	NT A	NT A	NT 4	NT 4	NT 4	NT A	NT 4		
GII ward	0	0	NA 1000/	NA 1000/	NA 0%	NA CC 70/	NA 1000/	NA (C70)	NA 1000/	NA	NA 0%	NA	NA 1000/	NA	NA 1000/	NA	NA 00/	NA	NA 1000/	NA		
Urology ward	2	3	100%	100%	0%	00.7%	100%	00.7%	100%	NA	0%	NA	100%	NA	100%	NA	0%	NA	100%	NA		
CIT ward	0	16	66 7%	100%	83 30%	100%	83 30%	66 7%	66 7%	00%	83 30%	70%	83 30%	50%	66 7%	87 5%	83 30%	100%	83 30%	66 7%		
Urology ward	3	63	100%	06.8%	65.570 66.7%	80%	33 3%	00.7% 85.7%	100%	90% 86.7%	65.3% 66.7%	7070 80%	03.370 33.30%	20%	100%	01.370	65.3% 66.7%	85 5%	33 30%	86.2%		
Pseudomonas aeruginosa	5	05	10070	90.070	00.770	8070	55.570	05.770	100%	80.770	00.770	8070	55.570	8070	100 %	94.070	00.770	05.570	55.570	80.270		
GIT ward	1	6	100%	75%	100%	NA	100%	0%	100%	100%	100%	NA	100%	50%	100%	NA	100%	NA	100%	NA		
Urology ward	2	32	100%	100%	100%	100%	100%	100%	100%	83.3%	100%	100%	100%	66.7%	100%	85%	100%	100%	100%	72%		
Kelebsiella pneumonia																						
GIT ward	0	3	NA	100%	NA	100%	NA	100%	NA	0%	NA	NA	NA	0%	NA	100%	NA	NA	NA	100%		
Urology ward	3	8	100%	75%	100%	100%	100%	50%	100%	100%	100%	NA	100%	0%	100%	100%	100%	100%	100%	100%		
Enterbacteriaceae sp																						
GIT ward	0	14	NA	100%	NA	0%	NA	100%	NA	NA	NA	NA	NA	NA	NA	65.3%	NA	55.6%	NA	77.8%		
Urology ward	5	42	100%	100%	100%		100%	100%	100%	NA	100%	NA	100%	NA	100%	89.8%	100%	94.4%	100%			
						100%														100%		
Non-lactose fermenting G-ve																						
GIT ward	0	3	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	100%	NA	100%	NA	NA		
Urology ward	0	11	NA	NA	NA	NA	NA	NA	NA	100%	NA	100%	NA	50%	NA	95.2%	NA	87.5%	NA	55.6%		
Serratia marcescens																						
GIT ward	0	0	NA	NA	NA	NA	NA	NA	NA	100%	NA	100%	NA	50%	NA	NA	NA	NA	NA	NA		
Urology ward	1	0	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA		
Proteus mirabilis	0	0	N T 4											N7 4								
GIT ward	0	0	NA	NA 1000	NA 1000/	NA	NA	NA	NA	NA	NA 1000/	NA 1000/	NA	NA 1000/	NA	NA	NA 1000/	NA	NA	NA		
Urology ward	2	5	/5%	100%	100%	NA	0%	0%	/5%	/5%	100%	100%	0%	100%	/5%	NA	100%	NA	0%	NA		

NA = Not Applicable (*) = decrease were significantly difference ($P \le 0.01$

Salah Ibrahim/ Elixir Pharmacy 41 (2011) 5657-5663

	Cyclic period (Rotation 2)																					
	No of	isolates		Cep	halospori	ns period (C	CEP)		Co-amoxiclave period (AMC)								Quinolones period (CIP)					
Organism (G+ve and																						
G-ve)			CEP (R%) AMC (R%		$CEP(R\%) \qquad AMC(R\%) \qquad CIP(R\%)$			(R %)	CEP(R%) AMC (R%)				CIP (R%)			CEP (P%) AM			$(\mathbf{R}\%)$ CIP (%)			
Patient location					ANIC	(R /0)	CII	(1(70)	CLI	(11/0)	AIVIC	(R /0)	Cli	(11/0)	CLI	(11/0)	AMC	(1(70)	CI	(70)		
		Before		Before	Before	After		Before		Before	Before	After		Before		Before		Before	Before	After		
		After		After	201010			After		After	201010	1 1101		After		After		After	201010			
Staphylococcus																						
aureus	14	13	97.6%	100%	100%	0%	72%	0%	97.6%	33.3%*	100%	66.7%	72%	NA	97.6%	77.8%	100%	87.5%	72%	$44.4\%^{*}$		
GIT ward	13	7		100%	100%	100%	92.3%	100%	100%	77.8%	100%	100%	92.3%	33.3%	100%	100%	100%	100%	92.3%	100%		
Urology ward			100%																			
Staphylococcus																						
saprophyticus	2	0	100%	NA	50%	NA	0%	NA	100%	NA	50%	NA	0%	NA	100%	NA	50%	NA	0%	NA		
GIT ward	2	0	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA		
Urology ward																						
Staphylococcus																						
epidermidis	1	0	100%	NA	NA	NA	0%	NA	100%	NA	NA	NA	0%	NA	100%	NA	NA	NA	0%	NA		
GIT ward	3	0	77.8%	NA	33.3%	NA	66.7%	NA	77.8%	NA	33.3%	NA	66.7%	NA	77.8%	NA	33.3%	NA	66.7%	NA		
Urology ward																						
Streptococcus sp.																						
GIT ward	0	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	100%	NA	0%	NA	0%		
Urology ward	0	7	NA	100%	NA	100%	NA	100%	NA	NA	NA	NA	NA	NA	NA	100%	NA	80%	NA	100%		
Entercoccus faecalis																						
GIT ward	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA		
Urology ward	2	0	100%	NA	0%	NA	100%	NA	100%	NA	0%	NA	100%	NA	100%	NA	0%	NA	100%	NA		
Escherichia coli																						
GIT ward	9	6	66.7%	NA	83.3%	NA	83.3%	NA	66.7%	100%	83.3%	75%	83.3%	$0\%^*$	66.7%	100%	83.3%	100%	83.3%	100%		
Urology ward	3	61	100%	100%	66.7%	95%	33.3%	95.2%*	100%	93.2%	66.7%	90.9%	33.3%	100%	100%	97.9%	66.7%	100%	33.3%			
25																				88%		
Pseudomonas																						
aeruginosa	1	1	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	100%	100%	100%	100%	100%		
GIT ward	2	36	100%	90.9%	100%	100%	100%	33.3%*	100%	92.9%	100%	NA	100%	42.9%	100%	60%	100%	NA	100%	30%		
Urology ward																						
Kelebsiella																						
pneumonia	0	1	NA	NA	NA	NA	NA	NA	NA	0%	NA	NA	NA	NA	NA	100%	NA	100%	NA	100%		
GIT ward	3	11	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	100%	80%	100%	100%	100%	100%	100%	100%		
Urology ward																						
Enterbacteriaceae sp																						
GIT ward	0	3	NA	NA	NA	NA	NA	NA	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%		
Urology ward	5	44	100%	100%	100%		100%	93.3%	100%	96.9%	100%	100%	100%	100%	100%	85%	100%	95%	100%			
						100%														83%*		
Non-lactose																						
fermenting G-ve	0	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	100%	NA	100%	NA	100%		
GIT ward	0	13	NA	100%	NA	100%	NA	50%	NA	100%	NA	100%	NA	NA	NA	100%	NA	100%	NA	72%		
Urology ward																						
Serratia marcescens																						
GIT ward	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA		
Urology ward	1	0	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA	100%	NA		
Proteus mirabilis																						
GIT ward	0	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	100%	NA	100%	NA	NA	NA	NA	NA	NA		
Urology ward	2	1	75%	100%	100%	100%	0%	0%	75%	NA	100%	NA	0%	NA	75%	NA	100%	NA	0%	NA		

NA = Not Applicable (*) = decrease were significantly difference (P \leq 0.01)