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# Preconcentration of trace Ni (II) using C<sub>18</sub> disks nano graphene with aminopropyltriethoxysilane (APTES)

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## ABSTRACT

A selective and simple method has been developed for preconcentration of Ni on  $C_{18}$  disks nano graphene with aminopropyltriethoxysilane (APTES). Simple chemical bonding method to synthesize nano graphene with aminopropyltriethoxysilane (APTES) was reported. The adsorption behaviors of Ni (II) in aqueous solution on APTES were systematically investigated. The procedure is based on the selective formation of Ni (II) at optimum pH by elution with organic eluents and determination by flame atomic absorption spectrometry. The method is based on complex formation on the surface of the ENVI-18 DISK<sup>TM</sup> disks nano graphene with amino propyltriethoxysilane (APTES) followed by stripping of the retained species by minimum amounts of appropriate organic solvents. The elution is efficient and quantitative. The effect of potential interfering ions, pH, APTES amount, stripping solvent, and sample flow rate were also investigated. Under the optimal experimental conditions, the break-through volume was found to about 1000mL providing a preconcentration factor of 600. The maximum capacity of the disk nano graphene with aminopropyltriethoxysilane (APTES) was found to be  $398\pm 3 \mu g$  for Ni <sup>2+</sup>. The limit of detection of the proposed method is 5ng per 1000mL.The method was applied to the extraction and recovery of Ni in different water samples.

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Introduction

Ni at trace concentrations acts as both a micronutrient and a toxicant in marine and fresh water systems (Izatt et al, 1991; Izatt et al,1985; Izatt et al,1995; Blake et al,1996; Arca et al,2001; Ghoulipour et al,2002; Hashemi et al,2001; Shcherbinina et al,1990). This element is needed by plants at only very low levels and is toxic at higher levels. At these levels, Ni can bind to the cell membrane and hinder the transport process through the cell wall. Ni at nearly 40ng mL<sup>-1</sup> is required for normal metabolism of many living organisms (Gomes-Gomes 1995; Unger et al, 1979). On the other hand, Ni is an important element in many industries. Thus, the development of new methods for selective separation, concentration and determination of it in sub-micro levels in different industrial, medicinal and environmental samples is of continuing interest. The determination of Ni is usually carried out by flame and graphite furnace atomic absorption spectrometry (AAS) . (Boudreau et al, 1989) as well as spectrometric methods (Bruening et al, 1991; Mahmoud and Soliman, 1997 a).

Solid phase extraction (SPE) methods are the best alternatives for traditional classic methods due to selective removal of trace amounts of metal ions from their matrices. Solid phase extraction determinations can be carried out on different efficient ways. One of the most appropriative performation features of SPE is achieved by using octadecyl silica membrane disks. SPE reduce the use of toxic solvent, disposal costs, and extraction time (Mahmoud et al, 1997 b,45; Mahmoud et al, 1997b).The octadecyl silica membrane disks involves shorter sample processing time and decreased plugging due to the large cross-sectional area of the disk and small pressure drop which allows higher flow-rates; reduced channeling resulting from the use of sorbent with smaller particle size and a greater mechanical stability of the sorbent bed(Tong et al,1990).

In our previous we modified SPE membrane disks with suitable for selective measurement of chromium(Dadler et al,1987; Moghimi 2007) and lead(Mahmoud et al,1990). Meanwhile, other investigators have successfully utilized these sorbents for separation and determination trace amounts of lead(Leyden et al,1976; Moghimi et al,2009; Liu et al,1992), copper (Liu et al,1996; Mishenina et al,1996; Wang et al,1999), silver( Wang et al,1997; Zhang et al,1982), mercury (Zhou et al,1983; Zargaran et al, 2008), cadmium (Tabarzadi et al, 2010), palladium (Shin et al, 2004), Ni (Moghimi et al,2012) and UO<sub>2</sub> (Mahmoud et al,1998; Moghimi et al,2006).

The  $\beta$ -cyclodextrin–chitosan nanoparticles have attracted attentions due to its many excellent properties such as easy separation and high adsorption capacity (Fang et al, 2007; Guo et al,2010) . In this work,  $\beta$  -cyclodextrin–chitosan/nano graphene oxide materials were prepared via a step procedure route that the carboxyl group of GO chemically reacts with the amine group of magnetic  $\beta$  -cyclodextrin–chitosan with consequent formation of chemical bond between GO and chitosan.

In the present report, we wish to describe a proper concentrative method for assessment of trace levels of Ni in different water samples. To the best of our knowledge, octadecyl silica membrane disks modified by (APTES) have not been used for Ni isolation and preconcentration, previously of this work was the development of a rapid, efficient and highly sensitive method for selective extraction and concentration of ultra trace amounts of Ni<sup>2+</sup> ions from aqueous media using octadecyl silica membrane disks nano graphene with aminopropyltriethoxysilane (APTES) and Flame Atomic Absorption Spectrometric (FAAS) determination.

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# Experimental

## Reagents

Graphite oxide was prepared from purified natural graphite (SP-1, Bay Carbon, Michigan, average particle size 30 lm) by the Hummers (Hummers et al, 1958). method and dried for a week over phosphorus pentoxide in a vacuum desiccators before use. 4-Isocyanatobenzenesulfonyl azide was prepared from 4carboxybenzenesulfonyl azide via a published procedure (Ibarra et al,1993). All solutions were prepared with doubly distilled deionized water from Merck (Darmstadt, Germany). C<sub>18</sub> powder for chromatography with diameter of about 50 µm obtained from Katayama Chemicals from supelco. It was conditioned before use by suspending in 4 M nitric acid for 20 min, and then washed two times with water. Sodium dodecvl sulfate (SDS) obtained from Merck (Darmstadt, Germany) and used without any further purification. The stock standard solution of Ni<sup>2+</sup> was prepared by dissolving 0.1000g of the Ni powder in 10mL concentrated nitric acid and diluted to 1000mL with water in a calibrated flask. Working solutions were prepared by appropriate dilution of the stock solution.

# Synthetic procedures

## **Preparation of GO–APTES**

Zhang et al. reported the preparation of chemical bonded graphene coating for SPME in a layer-by-layer manner (Fig. 1A) (Karousis et al,2011). The silica fiber was first treated with NaOH and 3- amino propyltriethoxysilane (APTES) to introduce amino groups to the surface. Then, GO was bonded to the fiber via reaction of the carboxyl groups of GO with the amino groups. Repeating the treatment with APTES and GO four times gave a coating of 20 lm thickness (Fig. 1D). Finally, the fiber was aged at 60°C and reduced with hydrazine. SEM showed a rough tree-bark-like structure with a striped appearance (Fig. 1B), and the high-resolution image revealed a continuous folded, wrinkled structure (Fig. 1D).



Figure 1. (A) Layer-by-layer fabrication processes of chemical-bonded graphene-coated SPE fiber. (B–D) SEM images of an SPME fiber coated with graphene. The surface images at magnifications of (B) 350· and (C) 5000·; the crosssection image at magnifications of (D) 3000(Karousis et al, 2011)

# Column preparation

GO–APTES (40 mg) were packed into an SPE mini-column (6.0 cm ×9 mm i.d., polypropylene). A polypropylene frit was placed at each end of the column to prevent loss of the adsorbent. Before use, 0.5 mol  $L^{-1}$  HNO<sub>3</sub> and DDW were passed through the column to clean it.

#### Apparatus

Determination of Ni<sup>2+</sup> contents in working samples were carried out by a Varian spectra A.200 model atomic absorption spectrometer equipped with a high- intensity hallow cathode lamp(HI-HCl) according to the recommendations of the manufacturers. These characteristics are tabulated in(Table 1).

GO–APTES (40 mg) were packed into an SPE ENVI-18 DISK<sup>TM</sup> disks. Solid phase extractions were carried out by glassy membrane disks, ENVI-18DISK<sup>TM</sup> 47mm diameter ×0.6 mm thickness containing octadecyl silica bonded phase (30  $\mu$ m particles, 70 A° pore size) obtained from Supelco in conjunction with a standard Millipore 47 mm filtration apparatus equipped with a vacuum pump. The pH measurements were carried out by an ATC pH- meter (EDT instruments, GP 353).

#### Sample extraction

Extraction were performed with glassy membrane disks, ENVI-18 DISK<sup>TM</sup> 47mm diameter ×0.6 mm thickness containing octadecyl silica bonded phase (30  $\mu$ m particles, 70 A° pore size) from Supelco. The disks were used in conjunctions with a standard Millipore 47mm filtration apparatus connected to water aspirator (Moghimi 2007).

1) Sample Treatment: The water samples were filtered through  $45\mu m$  nylon filters. Sampling vessels were polyethylene bottles soaked in 1 mol.L<sup>-1</sup> HNO<sub>3</sub> overnight and rinsed twice with deionized water. The analysis must be done within 2 days of sample collection to limit the risk of interconversion of Ni (II).Then, 5mL of methanol was added to a 90mL portion of each before analysis. The surface of the ENVI-18 DISK<sup>TM</sup> disks is modified with GO–APTES and therefore could retain Ni <sup>2+</sup> ions properly. Instead, 10 mg of GO–APTES an appropriate volume of an organic solvent (5mL) miscible with water. The most suitable solvent under the experimental conditions was acetone.

2) Disk cleaning and conditioning: A disk was placed in the apparatus and was washed with 10mL of methanol to remove all contaminants arising from the manufacturing process and the environment. Then, the disk was dried by passing air through it for several minutes. In order to insure optimal extraction of the analytes of interest, the disk was again washed with 10mL of methanol, immediately followed by 10mL of water, without letting the surface of the disk dry. This step pre-wets the disk surface before to extraction. Improper performance of this step causes slow flow – rate and poor analyte recoveries. It is important to avoid any air contact with the surface of the disk before the addition of the sample.

3) *Sample addition*: After complete homogenization, accurate volumes of the sample solutions (100mL portions) were transferred to the top reservoir of the disk apparatus. At the same time, the solution was drawn through the disk by applying a mild vacuum. Application of vacuum was continued until the disk was completely dry (about 5 minutes).

4)*Analyte elution* : In order to elute the analyte selectively, exactly 5 mL of acidified solvents 0.1M HCl in methanol was passed through the disk and collected into a 5.0 mL volumetric flask under the extraction funnel. It was found that ultra pure alcoholic organic solvents were the best eluting agents. The concentration of Ni (II) in the eluates were then determined by FAAS using an external calibration graph.

#### **Results and discussion**

The treatment of nano graphene with aminopropyltriethoxysilane (APTES) can lead to the derivatization of both the edge carboxyl and surface hydroxyl functional groups via formation of amides (Karousis et al,2011) or carbamate esters (Smith 2001), respectively.

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## Table 1. The operational conditions of flame for determination of Ni

Slit width	0.7 nm
Operation current of HI-HCL	15 mA
Resonance fine	324.8nm
Type of background correction	Deuterium lamp
Type of flame	Air/acetylene
Air flow	7.0 mL.min <sup>-1</sup>
Acetylene flow	1.7 mL.min <sup>-1</sup>

## Table 2. The effect of presence of GO-APTES on extraction percent of Ni (II)<sup>a</sup>.

GO-APTES	pН	Extraction percent of Ni (II)	
Absence	2-6	$0.03(7.3)^{b}$	
Presence	2-6	98.9(2.5) to 63(2.2)	
<sup>a</sup> Initial samples contained 10µg of Ni (II) in 100mL of water.			
<sup>b</sup> Values in parentheses are RSD <sub>s</sub> based on five individual replicate analyses.			

# Table 3.Effect of different eluting solvents on Percentage recovery of Ni (II) adsorbed on the disk <sup>a</sup>

		% Recovery	
Stripping solution	2mL	5mL	10mL
Methanol	82.3(2.6) <sup>b</sup>	92.5(2.7)	98.7(2.0)
Acidified methanol <sup>c</sup>	54.5(2.3)	83.5(2.6)	83.8(2.9)
Ammoniacal methanol <sup>d</sup>	53.4 (2.5)	87.5(2.6)	86.3(2.8)
Ethanol	82.5(1.3)	99.61.5)	99.0(2.3)
Acetonitril	36.6(4.3)	46.5(5.5)	69.5(2.9)
Formic acid(1M)10% V.V <sup>-1</sup> mthanol	55.2(1.2)	68.3(2.0)	71.5(2.8)
Hydrochloric acid(1M)10% V.V-1 mthanol	54.3(1.9)	92.0(2.6)	91.6(2.0)
Hydrochloric acid(1M)10% V.V <sup>-1</sup> mthanol	55.6(2.5)	65.3 (2.5)	97.7(1.6)
Nitric acid(2M)10% V.V <sup>-1</sup> mthanol	59.8(1.9)	85.9(2.3)	85.4(2.0)
Nitric acid(1M)10% V.V <sup>-1</sup> mthanol	64.0(2.5)	85.8(2.1)	87.7(1.6)
Ethanol	85.1(2.8)	95.7(2.0)	97.7(2.2)
<sup>a</sup> Initial samples contained 10 µg of each Ni in 100 mL water.			
<sup>b</sup> Values in parentheses are RSDs based on five individual replicate analysis.			
<sup>c</sup> Acidified solvents obtained by addition of $0.1M$ HCl			

<sup>d</sup>Ammoniacal solvents obtained by addition of 0.1M HC.

# Table 4. Percent recovery of Ni from the modified membrane disk in the presence of 0.01 M of different counter anions <sup>a</sup>

counter anion	%Recovery
Cl <sup>-</sup>	13.8
Br	23.9
ClO <sub>4</sub>	35.0
SCN	45.8
Picrate	76.9
Acetate	98.7
<sup>a</sup> Initial samples contain	ed 10ug of Ni (II) in 100mL of water.

## Table 5. Influence of the GO-APTES amount on the recovery of Ni (II) ions <sup>a</sup>

GO–APTES amount (mg)	Recovery(%) of Ni (II)	
2	34.25(2.6) <sup>b</sup>	
5	44.46(2.0)	
8	83.20(2.6)	
10	95.5(2.5)	
15	98.5(2.5)	
20	98.6(2.8)	
<sup>a</sup> Initial samples contained 10 µg of each Ni in 100 mL water.		
<sup>b</sup> Values in parentheses are RSDs based on five individual replicate analysis.		

## Table 6. Separation of Ni from binary mixtures <sup>a</sup>

Table 0. Separation of 10 monitorinary mixtures			
Diverse ion	Amounts taken(mg)	% Found	%Recovery of Ni <sup>2+</sup> ion
Na <sup>+</sup>	92.4	$1.15(2.4)^{b}$	98.5(2.7)
$\mathbf{K}^+$	92.5	1.36(2.3)	98.0(2.2)
Mg <sup>2+</sup>	24.5	0.70(2.6)	98.5(1.7)
Ca <sup>2+</sup>	26.3	2.65(3.0)	98.5(1.8)
Sr <sup>2+</sup>	2.45	2.85(2.1)	98.4(2.0)
Ba <sup>2+</sup>	3.66	3.16(2.1)	98.7(2.3)
Mn <sup>2+</sup>	2.66	1.75(2.2)	96.3(2.3)
Co <sup>2+</sup>	2.17	6.44(2.3)	93.0(1.9)
Cu <sup>2+</sup>	1.64	2.43(2.4)	93.7(2.4)
Zn <sup>2+</sup>	2.76	4.97(2.1)	97.6(2.4)
$Cd^{2+}$	2.77	2.96(2.4)	97.2(2.7)
Pb <sup>2+</sup>	1.74	2.74(1.9)	97.7(2.7)
$Hg^{2+}$	1.67	2.71(2.1)	97.7(2.7)
$Ag^+$	2.6i	3.47(2.9)	97.6(2.3)
UO <sup>2+</sup>	2.76	2.74(2.1)	98.3(2.7)
<sup>a</sup> Initial samples	contained 10µg Ni <sup>2+</sup> and differe	nt amounts of various	ions in 100 mL water(0.1 M acetate ion).
<sup>b</sup> Values in pare	ntheses are RSDs based on five	individual replicate ar	nalysis.

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Sample	Ni <sup>2+</sup> added ( $\mu$ g)	Ni <sup>2+</sup> determined(ng.mL <sup>-1</sup> )	ICP-AES
Tap water	0.0	1.72(2.8) <sup>a</sup>	ND
	10.0	11.93(2.0)	11.8
Snow water	0.0	4.85(2.5)	ND
	10.0	14.94(2.6)	14.5
Rain water	0.0	2.65(2.3)	ND
	10.0	12.85(2.4)	12.9
Sea Water	0.0	12.96(2.3)	12.8
	10.0	22.50(2.5)	23.0
<sup>a</sup> Values in parentheses are %RSDs based on five individual replicate analysis			
<sup>b</sup> Not detected.			

Table 7. Recovery of Ni added to 1000mL of different water samples (containing 0.1M acetate at pH= 4.0-4.5)

#### Morphology

The amount of nano graphene with aminopropyltriethoxysilane (APTES) sheet was evaluated by thermo-gravimetric analysis. As compared with the TGA results of pure graphite, which is thermally stable up to 900 °C under nitrogen, and GO which decomposes above 600 °C, after having lost the oxygenated species at 240 °C (i.e. 14.7% weight loss), the 6% weight loss occurred in the temperature range 250–550 °C for the GO–APTES material, is attributed to the decomposition of APTES (Fig. 2).



Fig. 2. The TGA graphs of graphite (black), GO (blue) and GO–APTES (red), obtained under an inert atmosphere.

The GO-APTES material forms a stable dispersion in DMF at a concentration not exceeding 1 mg mL<sup>-1</sup>. The electronic absorption spectrum of GO-APTES in DMF (Fig. 3), shows (i) a broad signal monotonically decreasing from the UV to the visible region, which is attributed to GO and (ii) a characteristic band at 420 nm (Soret-band) corresponding to the covalently grafted APTES units (the O-bands at 516, 557, 589 and 648 nm were flattened to the base line in the GO-APTES material). Interestingly, the absorption of porphyrin in the GO-APTES material is broadened, shortened and bathochromically shifted (ca. 2 nm) as compared to that of the free APTES, a result that corroborates not only the linkage of porphyrin with the GO sheets but also electronic interactions between the two species (i.e. GO and APTES) in the ground state. These results are in agreement with studies based on other hybrid systems consisting of porphyrins covalently grafted to carbon nanotubes and nanohorns(Karousis et al,2011).

#### **Evaluation of the role of the GO-APTES**

Some preliminary experiments were performed for investigation of absence or presence of GO–APTES on the quantitative extraction of Ni (II).It was concluded that the membrane disk itself does not show any tendency for the retention of Ni (II), but introduction of 100mL portions of aqueous Ni (II) samples containing  $10\mu g$  of Ni (II) and 10mg of GO–APTES leads to satisfactory its retention(Table 2). The latter case is most probably attributed to the existence of a considerable interaction between Ni (II) and the GO–APTES. It should be mentioned that formation of stable complexes between Ni (II) and GO–APTES at pH=2 is probably due to an ion pair formation mechanism. However, at pH higher than 2 the retention and percentage recovery of Ni (II) are negligible.



Fig 3. The UV-vis spectra of GO–APTES (black) and free APTES (red), obtained in DMF

#### Choice of eluent

In order to select the most appropriate eluent for the quantitative stripping of the retained Ni (II) on the disks, 5mL of various non organic (each containing 10% V.V<sup>-1</sup> methanol) and different organic solvents were tested. The results tabulated in Table2. As can be seen, the best eluting solvents were found to be 5mL of methanol or ethanol, resulting in quantitative elution of Ni (II) from the disk. It should be emphasized that presence of methanol in any kind of employed solvents helps to better the contact of eluent with hydrophobic surface of the disk.

#### Choice of pH

The pH of the sample solutions were adjusted to different values between 2-9 by addition of HCl or a buffer such as  $CH_3COONa$ -  $CH_3COOH$  or  $NaH_2PO_4$ , and then solutions passed through the sorbent.

Usually, the metal ions were stripped by pure methanol solutions followed by FAAS determination of the eluted Ni (II). Then, percentage recovery at various pH values was determined(Fig .4). According to the results shown in Fig.4 up to pH 4.0-4.5, complete percentage recoveries are obtained. However, at higher pH values, recovery decays. This is due to fact that in an acidic solution the protonation of GO-APTES occurs and there is a weak tendency for retention between Ni (II) and GO-APTES, whereas at higher values (pH>5), Ni (II) reacts with hydroxide ions to Ni (OH)<sub>2</sub>. Therefore, CH<sub>3</sub>COONa-CH<sub>3</sub>COOH buffer with pH=4.5 was used for the preconcentration step. Other solvents used for dissolving GO-APTES was methanol. The influences of these solvents on the percentage recoveries as a function of pH are compared and shown in Fig.4. Meanwhile, other organic solvents were not tested because of their restricted solubility and formation of two phases with aqueous solutions and incompatibility with flame higher pH values (>7) were not tested because of the possibility of the hydrolysis of  $C_{18}$  in the disks(Moghimi 2007). Ni (II) ions can be retained quantitatively by the modified membrane disk through the pH range from 4.0 to 4.5 However, at lower pH (< 4.0), nitrogen atoms of the GO–APTES could be protonated and the stability of complex is reduced.



Fig 4. Influence of sample pH on the percentage recovery of Ni (II)

# Effect amount of counter anion

In order to investigate the effect of counter ion on the recovery Ni <sup>2+</sup> ions by the modified disks, different counter anions were tested Table 4, it is immediately obvious that the nature of the counter anion strongly influences the retention of Ni <sup>2+</sup> ions by the disk. The results revealed that the GO–APTES behaves in pH range 4.0-4.5 (Leilei et al ,2013)so that the Ni <sup>2+</sup> ions are retained as ion pair complexes by the membrane disks. As seen, CH<sub>3</sub>COO<sup>-</sup> ion is the most efficient counter anion for the SPE of Ni <sup>2+</sup> ions. The influence of the concentration of CH<sub>3</sub>COONa ion on Ni recovery was investigated, and the results are shown in Table 4. As seen, the percent recovery of Ni <sup>2+</sup> increased with the CH<sub>3</sub>COO<sup>-</sup> concentration until a reagent concentration of about 0.1 M is reached, beyond which the recovery remained quantitative.

Moreover, acetate ion acts as a suitable buffering agent, while it effectively contributes to the ions- pair formation; thus, in the SPE experiments, there was no need for the addition of any buffer solution.



Fig 5. The effect of the flow-rate on extraction percent of Ni (II)

#### The influence of flow-rate

One of the most important parameters affecting solid phase extraction is the speed of the process. Hence, the effect of flow-rates on extraction efficiencies was investigated. It was found that in the range of 10-100 mL.min<sup>-1</sup>, the retention of Ni (II).was not considerably affected by the sample solutions flow-rates and leads to reproducible and satisfactory results (Fig. 5). Thus, the flow-rate was maintained at 89mL.min<sup>-1</sup> throughout the experiment.

#### **Quantity of the GO-APTES**

The optimum amount of GO–APTES for the quantitative extraction of Ni (II).was also investigated by adding various amounts of it to solution(between 2-20 mg). The results are listed in Table 5. The experimental results revealed that the extraction

of Ni (II).was quantitative using a sample solution containing more than 10 mg GO–APTES. Hence, subsequent extractions were performed with 15mg of GO–APTES.

## **Disk efficiency**

Undoubtedly, one of the major parameters affecting in the SPE determinations is the efficiency of the used membrane disks. However, to the best of our knowledge this case has not been discussed elsewhere in similar reports. Under the optimum experimental conditions, it was found out that each ENV-18 DISK<sup>TM</sup> disk could perform at least 14 replicate analyses if organic eluting solvents are used. On the other hand, acidic, eluents practically decrease the number of time a disk could be used to 10replicates. These observations are represented in Fig. 6.

#### **Analytical Performance**

When solutions of  $10\mu g$  Ni in 10, 50, 100, 600, 1000, 2000, 2500 and 3000mL solutions under optimal experimental conditions were passed through the disks, the Ni (II).was quantitatively retained in all cases. Thus, the breakthrough volume for the method must be greater than 2500mL, providing a concentration factor of >600. The limit of detection(LOD) of the method for the determination of Ni (II) was studied under the optimal experimental conditions. The LOD based on  $3\sigma$  of the blank(5mL of methanol) is 5 ng per 1000mL.

The capacity of modified disks(5mg GO–APTES ) was determined by passing 50mL portions of sample solutions containing 8mg of Ni and 0.1M sodium acetate-acetic acid buffer with pH 4.0-4.5, followed by the determination of the retained metal ions in the eluting solution using AAS. The maximal capacity of the disk obtained from three replicate measurements was  $398{\pm}3\mu g$  of Ni  $^{2+}$  on the disk.



Fig 6. Influence of eluent(5mL of methanol )type on disk efficiency

In order to investigate the selective separation and determination of Ni <sup>2+</sup> ions from its binary mixtures with various metal ions, an aliquot of aqueous solutions(50mL) containing 10µg Ni <sup>2+</sup> and mg amounts of other cations was taken and the recommended procedure was followed. The results are summarized in Table 6. The results show that the Ni (II) ions in binary mixtures are retained almost completely by the modified disk, even in the presence of up to about 100mg of various ions. Meanwhile, retention of other cations by the disk is very low and they can separated effectively from the Ni <sup>2+</sup> ion. Its is interesting to note that, in other experiments, we found that in the presence of high enough concentrations NH<sub>2</sub>OH.HCl as a suitable reducing agent(> 0.5M) (Moghimi 2007).

#### Analysis of water samples

In order to assess the applicability of the method to real samples, it was applied to the extraction and determination of Ni from different water samples. Tap water(Tehran, taken after 10 min operation of the tap),rain water(Tehran, 20January, 2013), Snow water (Tehran, 16 February ,2013)and Sea water(taken

from Caspian Sea, near the Mahmoud-Abad shore) samples were analyzed (Table 7). As can be seen from Table 4 the added Ni ions can be quantitatively recovered from the water samples used. As is seen, the recovered Ni ion reveals that the results are quite reliable and are in satisfactory agreement with those obtained by ICPAES.

## Conclusion

Results presented in this work demonstrate well the tremendous possibilities offered by the solid phase extraction of trace amounts of Ni (II) in water samples using Octadecyl Silica membrane disks modified by (GO–APTES) and its determination by FAAS. The developed method was simple, reliable, and precise for determining Ni in water. In addition, the proposed method was free of interference compared to conventional procedures to determine Ni (Moghimi et al,2012; Zargaran et al, 2008).The method can be successfully applied to the separation and determination of Ni in binary mixtures.

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## References

Ai L.H., Zhang C.Y., Chen Z.L. (2007), (Removal of methylene blue from aqueous solution by a solvothermal-synthesized graphene/magnetite composite), J. Hazard.Mater. 19: 4559–4563.

Arca M., Blake A.J., Casab J., Demartin F., Devillanova F.A., Garau A., Isaia F., Lippolis V., Kivekas R., Muns V., Schroder M., Verani G., (2001)( Conformationally locked pentadentate macrocycles containing the 1,10-phenanthroline unit. Synthesis and crystal structure of 5-oxa-2,8-dithia[9](2,9)-1,10phenanthrolinophane (L) and its coordination properties to Ni II, Pd II, Pt II, RhIII and Ru II ) J. Chem. Soc., Dalton Trans. 1180-1189.

Blake A.J., Demartin F., Deviallonova F.A., Garau A., Isaia F., Lippolis V., Schroder M., Verani G. (1996), (Synthesis of dinuclear group 6 metal carbonyl complexes bridged by 4,4'-bipyrazole ligands )J. Chem. Soc., Dalton Trans.3705-3712.

Boudreau S.P., Cooper W.T. (1989)(Analysis of thermally and chemically modified silica gels by heterogeneous gas-solid chromatography and infrared spectroscopy), Anal. Chem.61:41-47.

Bruening M.L., Mitchell D.M., Bradshaw J.S., Izatt R.M., Bruening R.L. (1991)(Removal of cesium from alkaline waste solution: Part II – Column ion exchange study), Anal. Chem. 63: 21-27.

Dadler V., Lindoy L.F., Sallin D., Schlaepfer C.W. (1987), (Solid Phase Extraction of Phenolic from Natural Water by Modified Polyacrylonitrile Fiber)Aust. J. Chem. 40 :1557-1563.

Deng H., Li X.L., Peng Q., Wang X., Chen J.P., Li Y.D. (2005), (Monodisperse magnetic single-crystal ferrite microspheres), Angew. Chem. Int. Ed. 44 :2782–2785.

Depan D., Girase B., Shah J.S., Misra R.D.K. (2011), Structure– process–property relationship of the polar graphene oxidemediated cellular response and stimulated growth of osteoblasts on hybrid chitosan network structure nanocomposite scaffolds, Acta Biomater. 7 :3432–3445.

Ghoulipour V., Husain S.W. (2002), (STUDIES ON TITANIUM(IV) TUNGSTOSILICATE AND TITANIUM(IV) TUNGSTOPHOSPHATE. II. SEPARATION AND ESTIMATION OF HEAVY METALS FROM AQUATIC ENVIRONMENTS) Acta Chromatogr.12: 170-174.

Fang, Z. Gu, D. Gang, C. Liu, E.S. Ilton, B. Deng, (2007), (Cr(VI) removal from aqueous

solution by activated carbon coated with quaternized poly(4-vinylpyridine)), Environ. Sci. Technol. 41:4748–4753.

Fan L., Luo C.N., Sun M., Li X.J. (2012), (Preparation of novel magnetic chitosan/graphene oxide composite as effective adsorbents toward methylene blue), Bioresour. Technol. 114 :703–706.

Gomes-Gomes M.M., Hidalgo Garcia M.M., Palacio Corvillo M.A. (1995) (On-line preconcentration of silver on a sulfhydryl cotton microcolumn and determination by flow injection atomic absorption spectrometry), Analyst120:1911-1916.

Guo Y.J., Guo S.J., Zhai Y.M., Dong S.J., Wang E.K., (2010) ( Photochemical Synthesis of Noble Metal (Ag, Pd, Au, Pt) on Graphene/ZnO Multihybrid Nanoarchitectures as Electrocatalysis for  $H_2O_2$  Reduction)ACS Nano 4 ;4001–4010.

Hashemi O. R ., Kargar-Razi M., Raoufi F., Moghimi A., Aghabozorg H., Ganjali M. R., (Ultra-trace monitoring of copper in environmental and biological samples by inductively coupled plasma atomic emission spectrometry after separation and preconcentration by using octadecyl silica membrane disks modified by a new schiff's base) (2001)Microchem. J.69:1-8.

Hummers W, Offeman R. Preparation of graphitic oxide. J Am Chem. Soc 1958;80:1339.

Ibarra L, Jorda C. Effect of a diazide as adhesive agent in elastomeric matrix-short polyamide fibers composite. J Appl Polym Sci 1993;48(3):375–81.

Izatt R.M., Bradshaw J.S., Nielsen S.A., Lamb J.D., Christensen J.J. (1985)( This Week's Citation Classic), D. Sen, Chem. Rev.85:271-277.

Izatt R.M., Pawlak K., Bradshaw J.S., Bruening R.L. (1991) (trans-Dichlorido(1,4,8,11-tetraazacyclotetradeca-

ne)manganese(III) tetrafluoridoborate), Chem. Rev.91:1721-1726.

Izatt R.M., Pawlak K., Bradshaw J.S., Bruening R.L. (1995) (Self-Assembled Ionophores. An Isoguanosine- $K^+$  Octamer), Chem. Rev.95:2529-2532.

Karousis N., Sandanayaka A. S. D., Hasobe T., Economopoulos S. P., Sarantopouloua E., Tagmatarchis N., J. Mater. Chem., 2011, 21:109-112.

Leyden D.E., Luttrell G.H., Nonidez W.K., Werho D.B. (1976)( Adsorption of Co(II) and Cu(II) on silica gel surface modified with pyridinium ion from acetone and ethanol solutions), Anal. Chem. 48:67-72.

Leilei L., Lulu F., Min S., Huamin Q., Xiangjun L., Huimin D., Chuannan L. (2013) (Adsorbent for chromium removal based on graphene oxide functionalized with magnetic cyclodextrinchitosan) Colloids and Surfaces B: Biointerfaces 107:76-83.

Liu Q.P., Liu J.C., Tong Y., Cheng J.K. (1992)(Development of a cloud-point extraction method for copper and nickel determination in food samples), Anal. Chim. Acta 269: 223-228.

Liu Q.P., Zhao T., Liu J.C., Cheng J.K. (1996) (E-T based statistical modeling and compact statistical circuit simulation methodologies), Microchim. Acta 122, 27.

Mahmoud M.E. (1997), (Silica gel-immobilized Eriochrome black-T as a potential solid phase extractor for zinc (II) and magnesium (II) from calcium (II)) Talanta 45: 309-314.

Mahmoud ME ,Soliman E.M. (1997 b)(Silica-immobilized formylsalicylic acid as a selective phase for the extraction of iron(III)), Talanta 44: 15-21.

Mahmoud M.E., Soliman E.M. (1997 a) (Study of the selective extraction of iron (III) by silica-immobilized 5-formyl-3-arylazo-salicylic acid derivatives), Talanta 44 :1063-1071.

Mahmoud M.E. (1999) (Selective solid phase extraction of mercury(II) by silica gel-immobilized-dithiocarbamate derivatives), Anal. Chim. Acta 398: 297-302.

Mahmoud M.E. (1998), in: Proceeding of the 25th FACSS Conference, Austin, TX, USA, 11–15 October.

Mishenina I.V., Shapovalova E.N., Bolshova T.A., Smirnov P.V., Shpigun O.A. (1996) (Comparison of the liquid and gas chromatography of five classes of metal complexes), J. Anal. Chem. 51: 270–276.

Moghimi, A. Ghiasi R., Abedin A.R., Ghammamy S. (2009) (Solid phase extraction of Cd(II) using mesoporous organosilicas and determination by FAAS. Afr. J. Pure Appl. Chem.3(3):051-059.

Moghimi, A. (2007), (Preconcentration and Determination of Trace Amounts of Heavy Metals in Water Samples Using Membrane Disk and Flame Atomic Absorption Spectrometry), Chinese Journal of Chemistry, 25(10): 640-645.

Moghimi A., Tehrani M.S., Waqif Husain S. (2006)( Preconcentration and Determination of Copper(II) Using Octadecyl Silica Membrane Disks Modified by 1,5-Diphenylcarhazide and Flame Atomic Absorption Spectrometry), Material Science Research India 3(1a): 27-32.

Moghimi , A .Abdouss M. (2012) (Preconcentration of Ni(II) from sample water by modified poly (ethylene terephthalate)-grafted-acrylic acid/acryl amide fiber), African Journal of Pure and Applied Chemistry , 6(8), 110 - 118.

Ramesh P., Bhagyalakshmi S., Sampath S. (2004), (Preparation and physicochemical and electrochemical characterization of exfoliated graphite oxide), J. Colloid Interface Sci. 274 :95–102.

Rana S., Rawat J., Misra R.D.K. (2005), (Anti-microbial active composite nanoparticles with magnetic core and photocatalytic shell:  $TiO_2$ -NiFe<sub>2</sub>O<sub>4</sub> biomaterial system, Acta Biomater. 1: 691–703.

Unger K., (1979)Porous Silica, Elsevier, Amsterdam.

Shcherbinina N.I., Myasoedova G.V., Khabazova T.A., Shroeva E.Y., Ishmiyarova G.R., Nikitina I.E., Bannykh L.N. (1990) (Preconcentration of trace amounts of silver ion in aqueous samples on octadecyl silica membrane disks modified with hexathia-18-crown-6 and its determination by atomic absorption spectrometry), Zh. Anal. Khim.45:2137-2141.

Shin D. H., Ko Y. G., Choi U. S., Kim W. N. (2004) (Fluorescence Enhancement of Coumarin Thiourea Derivatives

by  $Hg^{2+}$ ,  $Ag^+$ , and Silver Nanoparticles), Ind. Eng. Res. 43: 2060-2065.

Smith MB, March J. March's advanced organic chemistry: reactions, mechanisms, and structure. New York: John Wiley & Sons Inc.; 2001: 1182–3.

Tabarzadi M., Abdouss M., Hasani S. A., Shoushtary A.M., Mat.-wiss. u. (2010)( Ion adsorption studies of micro and nano acrylic fibers modified by ethanolamine. Ionen Adsorption von mit Ethanolamin modifizierten Mikro- und Nano-Acylfasern )Werkstofftech. 41(4):221-228.

Tong A., Akama Y., Tanaka S. (1990) (Sorption and preconcentration of some heavy metals by 2-mercaptobenzothiazole-clay). Anal. Chim. Acta 230: 179-185.

Vestal C.R., Zhang Z.J. (2003), (Synthesis and magnetic characterization of Mn and Co

spinel ferrite-silica nanoparticles with tunable magnetic core), Nano Lett. 3:1739-1743.

Wang H., Zhang H.S., Cheng J.K. (1999)( Studies on 2-(2-thiazolylazo)-5-diethylaminophenol as a precolumn derivatizing reagent in the separation of platinum group metals by high performance liquid chromatography), Talanta 48:1-8.

Wang H., Zhang H.S., Cheng J.K., Qiu P.H. (1997) ( APPLICATION OF HYPHENATED TECHNIQUES IN SPECIATION ANALYSIS OF ARSENIC, ANTIMONY AND THALLIUM) Microchem. J. 332-338.

Zargaran M., Shoushtari A. M., Abdouss M.(2008) (Ion adsorption studies of micro and nano acrylic fibers modified by ethanolamine. Ionen Adsorption von mit Ethanolamin modifizierten Mikro- und Nano-Acylfasern), J. Appl. Polym. Sci, 110:3843-3849.

Zhang C.P., Qi D.Y., Zhou T.Z. (1982)(Sensitive spectrophotometric determination of traces of zirconium with 2-(6-bromo-2-benzothiazolylazo)-5-diethylaminophenol in the presence of sodium lauryl sulphate), Talanta 29:1119-1123.

Zhou T.Z., Qi D.Y., Zhang C.P. (1983)( Development of a cloud-point extraction method for copper and nickeldetermination in food samples), Acta Chim. Sin. 41:237-242.