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Organic Chemistry



Synthesis, Characterization, Antibacterial Evaluation and Molecular Docking Studies of 2-Azetidinone Derivatives as Novel DNA Gyrase Inhibitors

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ABSTRACT

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Newly designed 2-azetidinone derivatives (4a-e) were synthesized in good yields and characterised by advanced spectroscopic studies. The title compounds were evaluated for qualitative (zone of inhibition) by agar diffusion method against Staphylococcus aureus, Bacillus subtilis, Escherichia coli, Pseudomonas aeruginosa and Streptococcus pyogenes. Some of the analogues were found to have comparable or even more potency than the standard drugs. Docking study was performed to check interaction of synthesized compounds with the target DNA gyrase.

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Keywords Hydrazone,

2-azetidinone, Antibacterial activity, Docking and DNA Gyrase.

Introduction

Increasing number of multi-drug resistant microbial pathogens and emerging infectious diseases are the major challenging problems for the clinicians. There is an urgent need to develop new class of potent antimicrobial agents. Heterocyclic compounds play an important role in designing of a new class of structural entities of medicinal importance with new mechanisms of action(Sumit Bansal et al. 2014). The most widely used antibiotics such as the penicillins, cephalosporins, carumonam, aztreonam, thienamycine and the nocardicins all contain β- lactam rings3. Biological activity of these heterocycles has helped the medicinal chemist to plan, organize and implement newer approaches towards the discovery of newer drugs. The most widely used antibiotics such as the penicillins, cephalosporins, carumonam, aztreonam, thienamycine and the nocardicins all contain β - lactam rings3. Biological activity of these heterocycles has helped the medicinal chemist to plan, organize and implement newer approaches towards the discovery of newer drugs (Kilarimath Basavaraj et al, 2014). In view of the general observation that pharmacological activity is invariably associated with a large variety of heterocyclic compounds, the investigation of some heterocycles such as azetidinone derivatives has been undertaken. 2-Azetidinones, commonly known as b-lactams are well-known heterocyclic compounds among organic and medicinal chemists. The 2-azetidinone derivatives have been reported to possess wide range of biological activities like antibacterial (Sharma et al., 1998), antifungal (Halve et al., anti-inflammatory (Gurupadayya et al., 2008), 2007). anticonvulsant (Rajasekaran and Murugesan, 2005). Recently, it has been reported that β -lactams have novel biological activities such as cytomegalovirus protease inhibitors(Kallappa M. Hosamani et al, 2009) and as inhibitors of acyl-CoA cholesterol acyltransferase (Rosenblum et al, 2000). The β -lactams also serve as synthons for many biologically important classes of organic Compounds(Jyotsna Meshram et al, 2010).

The present investigation deals with the clinically isolated different gram positive and gram negative bacteria against synthesized compounds and most of the tested compounds act as

tested compounds at active sites of protein receptors, topoisomerase II DNA gyrase enzymes (PDB ID. 2XCT) molecular docking studies were also preformed and reported in this article. DNA gyrase is a major bacterial protein that is involved in replication & transcription and catalyzes the negative supercoiling of bacterial circular DNA. The enzyme consists of two subunits. A and B, of molecular mass 97 and 90 kDa. respectively, with the active enzyme being an A2B2 complex. The A subunit of DNA gyrase is involved in DNA breakage and reunion while the B subunit catalyzes the hydrolysis of ATP(Amit Poshiya et al, 2011) The DNA gyrase is a known target for antibacterial agents since its blocking induces bacterial death. Hence, the studies are further extended to check probable interactions with this mostly preferred bacterial target(Smita pawar et al, 2014).

potent antibacterial agents. To understand the interactions of

The main objective of this paper is an approach of designing gyrase inhibitor molecules and their derivatives by docking simulations, followed by the synthesis, characterization and antibacterial evaluation was carried out.

Materials and methods

All the chemicals and solvents used were of AR grade obtained from Sigma Aldrich, Lobachemie (India). Melting points of the synthesized compounds were determined in openglass on a Staurt-SMP10 melting point apparatus and recorded in °0 C without correction. The purity of the compounds was ascertained by thin layer chromatography on silica gel coated aluminum plates (Merck) as adsorbent and UV light as visualizing agent. Synthesized compounds were recrystallised using ethanol as solvent. IR spectra were recorded on SHIMADZU FT-IR spectrometer using KBr pellet technique. ¹H-NMR spectra were recorded on BRUKER-400 spectrometer operating at 400 MHz using TMS as internal standard in DMSO (chemical shifts in ppm). Protein Structure Preparation The Xray crystal structures DNA gyrase (PDB: 3U2D) retrieved from the Research Collaboratory for Structural Bioinformatics (RCSB) Protein Data Bank (Figure 1) was used in this study.

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Water molecules of crystallization were detached from the composite and the protein was optimized for docking using the protein preparation and refinement utility provided by Schrödinger LLC. Partial atomic charges were assigned according to the OPLS-AA force field.

Results and discussions

Chemistry

General procedure for the synthesis of compounds 3a-e: In a 250 ml round bottom flask, a mixture of substituted carboxylic acid (0.1mol), ethanol (60 ml) and conc. H_2SO_4 (1.4 ml) were refluxed for 10 hours on a water bath. The solution was cooled and poured slowly with stirring on to 200 g of crushed ice. Sufficient ammonia solution was added to render the resulting solution alkaline, generally some ester separates as oil but most of it remains dissolved in the alkaline solution. The solution was extracted five times with ether (25 ml) the combined ethereal extract was dried with anhydrous MgSO₄. Ether was removed by evaporation on a water bath and the residue was collected. Physical data of ester was noted. The Synthetic procedure is shown in scheme 1.

General procedure for the synthesis of compounds $4a-e^{27}$: A mixture of ester and hydrazine hydrate in 1:1 portion and ethanol (30 ml) were taken in a round bottom flask and refluxed for 4-6 hrs. Excess of ethanol was removed by distillation. On cooling the product, acid hydrazide separates out. It was filtered and collected. Recrystallization was carried out with ethanol and physical data was noted. The physical properties of synthesized derivatives 3(a-e) are shown in Table 1.





Scheme I: Synthesis of the 2-Azetidinone derivatives 5(a-e) Synthesis of N'-(4-(difluoromethoxy)-3-hydroxybenzylidene) benzohydrazide (4a)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{15}H_{12}F_2N_2O_3$; M.Wt:306; yield 67%, mp 142-146 ⁰C; IR (KBr, m, cm ⁻¹): 2908 (-Ali CH), 3030 (-Aro CH), 1562 (-CH=N), 1687 (-C=O), 3197 (-OH) and 1172 (-C-F) ;1HNMR: (DMSO-d6, 400 MHz), δ 8.37 (s, 1H, CH=N), δ 7.12 (s, 1H, -NH), δ 6.93 (s, 1H, -OH), δ 7.30 (s, 1H, -CHF₂) and δ 7.12-7.92 (m, 8H, Aromatic protons)

Synthesis of N'-(4-(difluoromethoxy)-3-hydroxybenzylidene)-4-chlorobenzohydrazide (4b)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{15}H_{11}ClF_2N_2O_3$; M.Wt:340; yield 72%, mp 115-117 ⁰C; IR (KBr, m, cm⁻¹): 2960 (-Ali CH), 3134(-Aro CH), 1598 (-CH=N), 1637 (-C=O), 3317 (-OH), 1109 (-C-F) and 750 (-C-Cl);1HNMR: (DMSO-d6, 400 MHz), δ 8.30 (s, 1H, CH=N), δ 7.00 (s, 1H, -NH), δ 6.89 (s, 1H, -OH), δ 7.19 (s, 1H, -CHF₂) and δ 7.07-7.90 (m, 7H, Aromatic protons)

Synthesis of N'-(4-(difluoromethoxy)-3-hydroxybenzylidene)-4-nitrobenzohydrazide (4c)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{15}H_{11}F_2N_3O_5$; M.Wt:351; yield 65%, mp 110-112 ⁰C; IR (KBr, m, cm ⁻¹): 2931 (-Ali CH), 3062 (-Aro CH), 1589 (-CH=N), 1687 (-C=O), 3304 (-OH) and 1128 (-C-F; 1HNMR: (DMSO-d6, 400 MHz), δ 8.36 (s, 1H, CH=N), δ 7.04 (s, 1H, -NH), δ 6.94 (s, 1H, -OH), δ 7.23 (s, 1H, -CHF₂) and δ 7.04-8.15 (m, 7H, Aromatic protons)

Synthesis of N'-(4-(difluoromethoxy)-3-hydroxybenzylidene) isonicotinohydrazide (4d)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{14}H_{11}F_2N_3O_3$; M.Wt:307; yield 68%, mp 124-126 ⁰C; IR (KBr, m, cm ⁻¹): 2912 (-Ali CH), 3035 (-Aro CH), 1568 (-CH=N), 1645 (-C=O), 3288 (-OH) and 1170 (-C-F) ;1HNMR: (DMSO-d6, 400 MHz), δ 8.34 (s, 1H, CH=N), δ 7.11 (s, 1H, -NH), δ 6.93 (s, 1H, -OH), δ 7.30 (s, 1H, -CHF₂) and δ 7.14-8.77 (m, 7H, Aromatic protons)

Synthesis of N'-(4-(difluoromethoxy)-3-hydroxybenzylidene)-4-methoxybenzohydrazide (4e)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{16}H_{14}F_2N_2O_4$; M.Wt:336; yield 75%, mp 135-137 ⁰C; IR (KBr, m, cm ⁻¹): 2931 (-Ali CH), 3053 (-Aro CH), 1591 (-CH=N), 1687 (-C=O), 3329 (-OH) and 1130 (-C-F); 1HNMR: (DMSO-d6, 400 MHz), δ 8.33 (s, 1H, CH=N), δ 7.05 (s, 1H, -NH), δ 6.91 (s, 1H, -OH), δ 7.42 (s, 1H, -CHF₂), δ 3.83 (s, 3H, -OCH₃) and δ 7.04-7.91 (m, 7H, Aromatic protons)

General procedure for the synthesis of compounds (5a–e)

Hydrazones 3a-f (0.04 mol) and triethylamine (0.02 mol) in dioxan (20 mL) at $0-5^{\circ}$ C mixture was stirred for 5 h. During stirring, chloroacetyl chloride (0.01 mol) in dioxan (10 mL) was added dropwise. The mixture was refluxed for 2 h and kept for two days on room temperature. The resulting mixture was poured in the water and the solid was separated out. Recrystallization was done with ethanol–water or chloroform–water to give the azetidine-2-one, 4a-f compounds.

Synthesis of N-(3-chloro-2-(4-(difluoromethoxy)-3hydroxyphenyl)-4-oxoazetidin-1-yl)benzamide (5a)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{17}H_{13}ClF_2N_2O_4$; M.Wt:382; yield 64%, mp 208-210 ⁰C; IR (KBr, m, cm⁻¹): 2926 (-Ali CH), 3051 (-Aro CH), 1681 (-C=O), 3425(-OH); 1H NMR (400 MHz, DMSO-d6) (ppm): 4.83(d, 1H, -CH-N), 9.73 (d, 1H, OH), 4.53 (d, 1H, -CH-Cl) 7.09-8.04 (m, 4H, Ar-H)

Synthesis of 4-chloro-N-(3-chloro-2-(4-(difluoromethoxy)-3hydroxyphenyl)-4-oxoazetidin-1-yl)benzamide (5b)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{17}H_{12}C_{12}F_2N_2O_4$; M.Wt:417; yield : 60%, mp 235-237 ⁰C; IR (KBr, m, cm⁻¹): 2920 (-Ali CH), 3047 (-Aro CH), 1664 (-C=O), 3435 (-OH); 1H NMR (400 MHz, DMSO-d6) (ppm): 4.62(d, 1H, -CH-N), 9.89(s, 1H, OH), 4.48(d, 1H, -CH-Cl) 6.86-7.82(m, 4H, Ar-H)

Synthesis of N-(3-chloro-2-(4-(difluoromethoxy)-3hydroxyphenyl)-4-oxoazetidin-1-yl)-4-nitrobenzamide (5c)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{17}H_{12}ClF_2N_3O_6$; M.Wt:427; yield : 56%, mp 240-242 ⁰C; IR (KBr, m, cm ⁻¹): 2926 (-Ali CH), 3051 (-Aro CH), 1664 (-C=O), 3431 (-OH); 1H NMR (400 MHz, DMSO-d6) (ppm): 4.37(d, 1H, -CH-N), 11.89 (s, 1H, OH), 4.14 (d, 1H, -CH-Cl) 7.16-8.35 (m, 4H, Ar-H)

Synthesis of N-(3-chloro-2-(4-(difluoromethoxy)-3hydroxyphenyl)-4-oxoazetidin-1-yl)isonicotinamide (5d)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{16}H_{12}ClF_2N_3O_4$; M.Wt:383; yield : 65%, mp 210-212 ⁰C; IR (KBr, m, cm⁻¹): 2924 (-Ali CH), 3045 (-Aro CH), 1674 (-C=O), 3425 (-OH); 1H NMR (400 MHz, DMSO-d6) (ppm): 4.77 (d, 1H, -CH-N), 11.84 (s, 1H, OH), 4.72 (d, 1H, -CH-Cl) 6.56-8.35 (m, 4H, Ar-H)

Synthesis of N-(3-chloro-2-(4-(difluoromethoxy)-3hydroxyphenyl)-4-oxoazetidin-1-yl)-4-methoxybenzamide (5e)

This was prepared and purified as per the above mentioned procedure: M.F: $C_{18}H_{15}ClF_2N_2O_5$; M.Wt:412; yield : 60%, mp 182-184 ⁰C; IR (KBr, m, cm⁻¹): 2927 (-Ali CH), 3043 (-Aro CH), 1672 (-C=O), 3425 (-OH); 1H NMR (400 MHz, DMSO-d6) (ppm): 4.78(d, 1H, -CH-N), 11.69 (s, 1H, OH), 4.69 (d, 1H, -CH-Cl) 6.91-7.91 (m, 4H, Ar-H)

MOLECULAR DOCKING STUDIES

Ligand Structure Preparation

The ligand structures were constructed using the splinter dictionary of Maestro 9.3 (Schrodinger, LLC) using the Optimized Potentials for Liquid Simulations-All Atom (OPLS-AA) force field with the steepest descent followed by curtailed Newton conjugate gradient protocol. Partial tomic charges were computed using the OPLS-AA force field (Pandurangan Perumal).

Docking Protocol

All docking calculations were performed using the "Extra Precision'' (XP) mode of GLIDE program. The binding site, for which the various energy grids were designed and stored, was defined in terms of two concentric cubes: the bounding box, which must contain the center of any acceptable ligand pose and enclosing box. It must include all ligand atoms of an acceptable pose, with a Root Mean Square Deviation (RMSD) smaller amount than 0.5 Å and an utmost atomic dislocation of less than 1.3 Å were eliminated as an outmoded in order to increase diversity in the retained ligand poses. The scale factor for van der Waals radii was applied to those atoms with absolute partial charges less than or unchanged to 0.15 (scale factor of 0.8) and 0.25 (scale factor of 1.0) electrons for ligand and protein, correspondingly. The max keep variable which sets the maximum number of poses generated during the initial phase docking calculation were set to 5000 and the keep best variable which sets the number of poses per ligand that enters the energy minimization was set to 1000



Figure 1. X-ray crystal structure of protein DNA gyrase (PDB: 3U2D)

Energy minimization protocol includes dielectric constant of 4.0 and 1000 conjugate gradient steps. Upon completion of each docking calculation, at most 100 poses per ligand were generated. The paramount docked structure was chosen using a GLIDE score (Gscore) function. One more scoring function used by GLIDE E-model, which itself consequent from a combination of the Gscore, Columbic, van der Waals and the strain energy of the ligand.

Qikprop Analysis

Qikprop efficiently evaluates pharmaceutically relevant properties for over half a million compounds per hour, making it a vital lead creation and lead optimization implement. Accurate prediction of Absorption, Distribution, Metabolism, Elimination (ADME) properties prior to expensive experimental procedures, such as High Throughput Screening (HTS), can eliminate unnecessary testing on compounds that will ultimately fail; ADME prediction can also be used to focus lead optimization efforts to enhance the desired properties of a given compound.

Considering the well obtained in vitro results, it was thought worthy to perform molecular docking studies, hence screening the compounds, inculcating both in silico and in vitro results. Considering DNA gyrase as the target receptor, comparative and automated docking studies with newly synthesized compounds was performed to determine the best in silico conformation. Fig. 1 shows the native crystal structure of DNA gyrase was obtained from Protein Data Bank (http://www.pdb.org/pdb/home/home.do) with the PDB ID 3U2D.

The active pocket consisted of amino acid residues as LEU 103, VAL 131, 79 & 130, ILE 51, 86, 102 & 175, THR 173, SER 55, & 129, ASP 81, ASN 54, GLU 58, GLY 83 & 85, PRO 87 and ARG 84 & 144 as shown in Fig. 3. The synthesized ligand molecules having 2D structure were converted to energy minimized 3D structures and were further used for in silico protein–ligand docking. All the five synthesized molecules (5a-e) were docked. Fig.2 shows the docked images of selected ligands including the considered standard drug i.e. ciprofloxacin.



Figure 2: Docking of synthesized azetidinone derivatives (5ae) with reference standard ciprofloxacin into the active site of DNA gyrase. The amino acids involved in hydrogen, hydrophobic and van der Waals interactions are highlighted.

Table 2 shows the Binding Energy and glide score of six compounds including the standard. In silico studies revealed all the synthesized molecules showed good binding energy toward the target protein ranging from Derivatives 5a, 5b and 5e shows hydrophobic intractions with the active site of gyrase (Fig. 3). Energy scores (S) and docking scores for compounds 5a, 5b, 5c, 5d and 5e were listed in Table (2). Compound 5c revealed docking score -6.018386 with energy score (S) -45.487493kcal/mol and interacted with ARG 84 with hydrogen bonds through NO₂ of the phenyl moiety (Fig. 3). Compound 5d showed the docking score -6.064761 with energy score (S) -43.744902kcal/mol and interacted with ASN 54 with a hydrogen bond through a N-H of the Azetidinone ring (Fig.3). Apart from hydrogen bonding, hydrophobic and van der Waals interactions were also detected between lead molecule 5a, 5b and 5e with DNA gyrase. The entire synthesized azetidinone derivatives were projected into the hydrophobic pocket of DNA gyrase defined by LEU 103, VAL 131, 79 & 130, ILE 51, 86, 102 & 175, THR 173, SER 55, & 129, ASP 81, ASN 54, GLU 58, GLY 83 & 85, PRO 87 and ARG 84 & 144 amino acid residues of the target protein (Fig.3).

From the docking study we predicted that Azetidinone analogues (5a, 5b, 5c, 5d and 5e) possess better antibacterial activity by having good binding affinity with target protein and it could be used as potential drugs as antimicrobial. Amongst all the docked compounds, the compound (5b) shows good binding affinity & interaction docking score of -6.4676 with DNA gyrase enzymes with reference to ciprofloxacin shown in figure 3.



Figure 3. Binding conformation and interaction of ligands 5a, 5b, 5c, 5d 5e and standard ciprofloxacin within the active site of DNA Gyrase

5e

21

21

Antibacterial Activity

All the synthesized compounds were evaluated *in vitro* for antibacterial activity by using filter paper disc method(E.W. Koneman et al, 1997; V. Cleidson et al, 2007) against different

strains of bacteria viz. B. substilis, E. coli, S. aureus Pseudomonas aeruginosa and Streptococcus pyogenes. All the compounds along with standard antibacterial Ciprofloxacin were used at 150µg concentrations. Nutrient agar medium was used for the study. After sterilization the nutrient agar medium was melted, cooled and inoculated with three Gram-positive organisms S. aureus, B. subtilis, Streptococcus pyogenes and two Gram-negative organisms E. coli, P. aeruginosa and poured into sterile Petri dish to get a uniform thickness of 6 mm. Dried and sterilized filter paper discs (6mm in diameter) soaked with known amount of test agents were placed on the nutrient agar media solidified in petridishes .The standard antibacterial agent Ciprofloxacin (150 µg), solvent control and the synthesized compounds in a concentration of 150 µg were The plates were then incubated at 37°C for 24 h and the diameter of zone of inhibition were measured and recorded in Table 3.

The zone of inhibition of all compounds is presented in Table 1 and Figure 8. Ciprofloxacin was used as standard drug for comparison with test compounds. All the tested compounds showed good antibacterial activity against Gram-positive and gram-negative strains. Among tested compounds, 5b showed significant zone at 16mm and 15mm against B. subtilis and S. aureus, respectively and found moderate active against both Gram-negative strains E. coli (13mm) and P. aeruginosa(16mm). Compound possessing electron withdrawing chloro group (5a) showed moderate activity against both Gram-positive bacteria when compared to other derivatives in the series (Table 3).All other derivatives shown moderate to good activity against bacterial strains.



Figure 4. In vitro antibacterial activity of synthesized azetidinone compounds 5a-e

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Compound	R	Molecular	M.Pt	M.Formula	Physical	IR Frequency
-		Tormula	0		state	
3a	C_6H_5	$C_7H_8N_2O$	110	136	White	3032 cm^{-1} (C-H Ar str); 1622 cm ⁻¹ (C=O str); 3296, 3215
					powder	$(NHNH_2 \text{ str}); 985 \text{ cm}^{-1} (N-N \text{ str}).$
3b	C ₆ H ₅ Cl	C ₇ H ₇ ClN ₂ O	162	170	White	3016 cm ⁻¹ (C-H Ar str); 1645 cm ⁻¹ (C=O str); 3302, 3209 cm ⁻¹
					powder	$(NHNH_2 \text{ str}); 727 \text{ cm}^{-1} (C-Cl \text{ str}); 987 \text{ cm}^{-1} (N-N \text{ str}).$
3c	C ₆ H ₅ NO ₂	$C_7H_7N_3O_3$	215	181	Pale Yellow	3070 cm ⁻¹ (C-H Ar str); 1662 cm ⁻¹ (C=O str); 3296, 3180 cm ⁻¹
					powder	(NHNH ₂ str); 1525 cm ⁻¹ (NO ₂ str); 1055 cm ⁻¹ (N-N str).
3d	C ₅ H ₅ N	C ₆ H ₇ N ₃ O	170	137	White	3064 cm ⁻¹ (C-H Ar str); 1649 cm ⁻¹ (C=O str); 3305, 3178 cm ⁻¹
					powder	$(NHNH_2 \text{ str}); 1045 \text{ cm}^{-1} (N-N \text{ str}).$
3e	C ₆ H ₅ OCH ₃	$C_8H_{10}N_2O_2$	135	166	White	3043 cm ⁻¹ (C-H Ar str); 2933 cm ⁻¹ (C-H Aliphatic str); 1612
					powder	cm^{-1} (C=O str); 3381, 3253 cm^{-1} (NHNH ₂ str); 1031 cm^{-1} (N-
						N str).

 Table 1. Physical properties and IR Spectral Data of Synthesized Hydrazides 3(a-e)

Table 2. Molecular docking studies of synthesized 2-Azetidinone Derivatives 5(a-e)compounds

S.No	Compound	Glide Score	Glide Energy
1	5a	-5.918678	-39.383112
2	5b	-6.467631	-42.172485
3	5c	-6.018386	-45.487493
4	5d	-6.064761	-43.744902
5	5e	-6.396769	-40.713267
7	Ciprofloxacin	-7.636035	-30.0573

Table 5. Antibacterial activit	Table 3.	Antibacterial	activity
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		Standard Antibiotic Disk*	Zone of inhibition mm in diameter				
S. No.	Bacteria		1	2	3	4	5
			5a	5b	5c	5d	5e
1	Bacillus subtilis	24	18	16	16	15	12
2	Escherichia coli	18	10	13	08	12	10
3	Pseudomonas aeruginosa	16	13	16	13	14	14
4	Staphylococcus aureus	18	13	15	12	13	12
5	Streptococcus pyogenes	28	12	16	14	15	16

Conclusion

A series substituted N-(3-chloro-2-(4-(difluoromethoxy)-3hydroxyphenyl)-4-oxoazetidin-1-yl) derivatives benzamide (5ae) were synthesized in good yields and were evaluated for antibacterial activities. Docking studies has been carried out by using Maestro 9.3 software for all the derivatives. The structures of the synthesized compounds were confirmed by FT-IR, 1H NMR spectral analysis. All the derivatives of 5a-e were docked with target protein for DNA Gyrase to predict the antibacterial activity. The results of the docking study for all the derivatives are shown in table. Amongst all the docked compounds, the compound (5b) shows good binding affinity & interaction docking score of -6.4676 with DNA gyrase enzymes with reference to ciprofloxacin. The pharmacological study was undertaken to evaluate the effects of substituents on the antibacterial activity. All the synthesized compounds exhibited good antibacterial activity towards Gram-positive bacteria and gram negative bacteria strains. The theoretical results obtained in the molecular docking were compared with the experimental results. The theoretical results obtained after docking of azetidinone analogues with DNA gyrace showed good agreement with the experimental results. Thus it confirms that, the experimental values moderately agree with theoretical values, which suggest that the parameters for docking simulation are optimum in reproducing experimental orientation of these compounds.

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