

Comparative Study on the antibacterial activity in the coelomic fluid of *Eudrilus eugeniae* (Kinberg, 1867) and *Polypheretima elongata* (Perrier, 1872)

P.K.Ramasamy¹, C.Murthy², V.Gokula¹, K.Govindaraj¹ and T.Manikandan¹

¹Department of Zoology, National College (Autonomous), Tiruchirappalli- 620001. Tamil Nadu, India.

²Department of Zoology & Biotechnology, AVVM Pushpam College (Autonomous), Poondi, Thanjavur-613503 Tamil Nadu, India.

ARTICLE INFO

Article history:

Received: 28 January 2017;

Received in revised form:
17 March 2017;

Accepted: 28 March 2017;

Keywords

Antibacterial activity,
Coelomic fluid,
Eudrilus eugeniae,
Polypheretima elongata,
Staphylococcus aureus,
Pseudomonas aureus and
E.coli

ABSTRACT

In the present investigation an attempt was made to test the antibacterial activity in the coelomic fluid of the epigeic species of earthworm, *Eudrilus eugeniae* and the anecic species of earthworm, *Polypheretima elongata*. The *Eudrilus eugeniae* was collected from Selvam Vermicomposting farm, Udumalpet, Tamil Nadu, India. Likewise, the *Polypheretima elongata* was also collected from the soil of National college campus, Tiruchirappalli, Tamil Nadu, India. Both worms were identified by following standard taxonomical key characters. After identification, both worms were cultured under favorable laboratory condition for 30 days providing the food material consisting of 50% predecomposed organic waste and 50% predecomposed cow dung in separate cement tanks. At the end of 30 days a pool of 20 healthy worms was selected from both cultures for the extraction of coelomic fluid. Later on, the fluid was extracted by electric shock method employing 9 volt electrode. The extracted coelomic fluid (100%) was used to test the antibacterial activity against two gram positive bacteria namely *Staphylococcus aureus* and *Pseudomonas aeruginosa* and one gram negative bacterium namely *E.coli*. Among the selected bacterial strains, the coelomic fluid of *Eudrilus eugeniae* and *Polypheretima elongata* showed the maximum inhibition zones 7.01 ± 0.28 mm and 2.43 ± 0.38 mm against *Staphylococcus aureus* respectively when compared to inhibition zones 2.35 ± 0.31 mm and 2.28 ± 0.29 mm; 2.66 ± 0.31 mm and 1.35 ± 0.17 mm produced by *Eudrilus eugeniae* and *Polypheretima elongata* against *Pseudomonas aeruginosa* and *E.coli* respectively. Besides that the coelomic fluid of *Eudrilus eugeniae* exhibited more inhibition activity than the zone (3.70 ± 0.13 mm) produced by the positive control (standard antibiotic, penicillin) against *Staphylococcus aureus*. But at the same time there is no remarkable difference between the inhibition zone of the coelomic fluid of *Polypheretima elongata* and the positive control experiment against all the bacterial strains tested. However, the control experiment showed more inhibition zone (3.97 ± 0.24 mm) than the inhibition zone produced by the coelomic fluid of *Polypheretima elongata* against *Pseudomonas aeruginosa*. As for the present study the bacterium *Staphylococcus aureus* activity is highly inhibited by the coelomic fluid of both species of earthworms than the other bacterial strains tested even though there is remarkable difference in the size of zone of inhibition. The data were statistically analyzed at 5.0% level employing ANOVA.

© 2017 Elixir All rights reserved.

Introduction

Earthworms are macro invertebrates which constitute the most important group of the soil invertebrates biodiversity in terms of their influence on soil formation and maintenance of soil fertility in agro-ecosystems (Edward *et al.*, 1995), and also form more than 80% of the invertebrate biomass in world ecosystems (Senapati, 1992). Earthworms are physically aerators, crushers and mixers; chemically decomposers; and biologically stimulators, in the decomposer systems. They effectively harness the beneficial microflora, destroy the soil pathogens and convert the organic wastes into vitamins, enzymes, antibiotics, growth hormones and protein rich casts (Prabha *et al.*, 2007). Of late, these

organisms have drawn the attention of scientists because of their involvement in organic waste degradation to produce a marketable bio-fertilizer, vermicompost (Edwards and Bohlen, 1996). It is a good substitute for commercial fertilizers and has high level of N, P and K than the normal heap manure (Srivastava and Beohar, 2004). Besides, these organisms are a source of animal protein in the diet of animals such as poultry birds and fish (Harwood, 1976; Mekada *et al.*, 1979; Sabine, 1978; Guerrero, 1983; Xu *et al.*, 1984; Stafford and Tacon, 1988) and indigenous people (Edwards and Lofty, 1972; Macdonald, 1983). Apart from this, recently earthworms play an important role in the pharmacological studies.

Tele:

E-mail address: kadur_rama@yahoo.co.in

© 2017 Elixir All rights reserved

Earthworms have been used in medicine for various remedies since 1340 AD (Stephenson, 1930). Earthworm has been recognized in oriental medicine as anti-inflammatory, analgesic and anti-pyretic agent (Noda *et al.*, 1992). It shows anti-cancer effect by preventing excess glucose uptake (Nagasawa *et al.*, 1992). Microorganisms are known to play a major role in soil characteristics; invertebrates are believed to act as regulators of antimicrobial activity. Earthworm surface excreta were found to have potent antimicrobial activity (Oleynik and Byzov, 2008).

It is also having anti-coagulatory and fibrinolytic activity which results in the facilitation of blood circulation (Wang *et al.*, 1989). The earthworm has been suspected to contain proteases which dissolve the fibrin clots are anticoagulant which selectively interfere with the intrinsic pathway of blood coagulation cascade (Mannet *et al.*, 1990 Davie *et al.*, 1991, Leipner *et al.*, 1993 Kim *et al.*, 1995 and Woo *et al.*, 1996). Medicinal properties of earthworm have also been described by authors like Bristow (1932), Ogata and Mori (1938), Ogata *et al.* (1939), Carr (1951) and Yegnanarayan *et al.* (1987).

Besides that earthworms have largely been used internally and externally as powerful aphrodisiacs (Vohra and Khan, 1978). The anti-inflammatory and anti-pyretic activities of biologically isolated extract from whole earthworm, *Lampito mauritii* was determined (Balamurugan *et al.*, 2008). Antimicrobial potency of *Eudrilus eugeniae* extracts on certain plant pathogens was studied (Shobha and Kale, 2007). Several isolated bacteria strains exhibited a high pathogenic effect when injected into *Eisenia coelomic* cavity (Lassbagues *et al.*, 1981). Anti-tumor activities of earthworm fibrinolytic enzyme on human hepatoma cells were studied (Hong, 2007). In 1983 it was also reported that very strong and novel fibrinolytic enzymes could be extracted from the earthworm, *Lumbricus rubellus*. These enzymes were fractionated and purified as six novel fibrinolytic enzymes and named collectively as Lumbrokinase (Mihra *et al.*, 1996; Tang *et al.*, 2000; Li *et al.*, 2008). It was also found that earth worm powder contains two kinds of inhibitory substance for the platelet aggregation induced by collagen and ADP. This novel substance also displays a relaxation effect for the canine saphenous vein induced by prostaglandin F in vitro and an inhibitory effect on the active partial Thromboplastin time (APTT) (Mihara *et al.*, 1996). Earthworm powder can be given orally; thus for this reason, earthworm powder has potential application as a thrombolytic and also exerts an inhibitory effect on platelet aggregation, an anticoagulant effect and a relaxation effect for the vascular systems, which are all effective for thrombotic therapy (Kim *et al.*, 1998). Earthworm powder represents a very promising agent for the treatment of thrombosis (Mihara *et al.*, 1996). Its tonic properties also make it beneficial support for the live and other organ system.

In china, Korea, Vietnam and most of Southeast Asia, *Lumbricus* has been used for their therapeutic benefits for thousands of years and referred to as earth dragons. It has been found that coelomic fluid of the earthworm contain more than 40 proteins and exhibits several biological activities as follows; cytolytic, proteolytic, antimicrobial, hemolytic, hemagglutinating, tumorolytic, mitogenic activities (Cotuk and Dales 1984; Lange *et al.* 1997; Lange *et al.* 1999; Cooper and Roach 2003; Lassagues *et al.*, 1981) found that this hemolytic factor also inhibited the growth of

different bacterial species which were isolated from nature. The coelomic fluid of *Eisenia foetida* Andrei was demonstrated to possess an antimicrobial activity against *Aeromonas hydrophila* and *Bacillus megaterium* which are known as earthworm pathogens (Valembouis *et al.* 1982 and Pan *et al.* 2003). Afterwards, Milocha *et al.* (1997) obtained two proteins, named Fetidins from dialyzed coelomic fluid of earthworms and confirmed that this antibacterial activity was due to fetidins. Cho *et al.* (1998) found that *Lumbricus rubellus* also has two antibacterial agents named Lumbricin 1 and Lumbricin 2. Of late, Kathireswari *et al.* (2014) have reported that the antimicrobial activity of the two species of earthworms namely *Lampito mauritii* and *Megascolex konkanensis* have revealed the maximum inhibition activity against the bacterial strains such as *Aeromonas hydrophilla*, *Bacillus subtilis* and *Vibrio para haemolyticus* and they also stated that the coelomic fluid of the earthworms namely *Darwida impertusa* and *Darwida lennora* showed less inhibition activity against these bacterial strains.

Recently, two types of antibacterial factors which include lysozyme-like molecules with hemolytic activity as well as a pattern recognition protein named coelomic cytolytic factor (CCF) have been identified in *Eisenia foetida* earthworm (Kohlerova *et al.* 2004). Bruhn *et al.* (2006) started that lysenin which was a different protein of *Eisenia foetida* and lysenin-like proteins had several cytolytic activities which exerted hemolytic, antibacterial and membrane-permeabilizing properties. Many proteins are involved in immunodefense of earthworms, such as lysozyme, lysenin, fetidins and CCF-1 (Cooper *et al.* 2002). However, it is not known whether antimicrobial peptides contribute to the innate defense of earthworms. To date only one antimicrobial peptide, lumbricin I, has been isolated from the earthworm *Lumbricus rubellus*. Lumbricin I, which belongs to proline- rich antimicrobial peptides group, kills both bacteria and fungi without hemolytic activity in vitro (Cho *et al.* 1998). Despite their immense economic importance, the investigation on the antibacterial activity of earthworms is limited to three to four species. Hence, in the present investigation an attempt was made to bring out new antimicrobial agent or components from the coelomic fluid of earthworms, *Eudrilus eugeniae* and *Polypheretima elongata*. In addition to that the present study will definitely lead to formulation of new natural antimicrobial agent and this may be found useful in future prospectus for mankind.

Materials and methods

Collection and identification of earthworms

The earthworms were collected with the help of shovel by digging and hand sorting method from the soil of National College Campus Area. The soil was dug up to a depth of 0-100 cm and the worms were hand collected cautiously without any damage and they were washed in clean surface water. Among the collected worms some of them were segregated to study the taxonomical characters. They were allowed in enamel tray half filled with water. They were narcotized by gradually adding ethanol to water. The narcotized worms were fixed in 5%-10% formalin solution depending on the size of the worms for 24 hrs in straightened position. The fixed worms were examined morphologically and anatomically to study the taxonomical characters (Julka and Paliwal, 1993).

Both external and internal characters are considered important in earthworm taxonomy. It is therefore, necessary to dissect the earthworms for their internal observations.

Earthworms were examined in various growth stages. This is indicated in the material examined as 2-4-7, denoting 2 juveniles, 4 acitellate and 7 clitellate worms (Julka, 1988). Lengths of uniformly contracted worms were measured with a linear scale, and their diameter was measured on segments just anterior to clitellum. The worms were dissected longitudinally slightly left or right side of the mid-dorsal line by using a sharp tissue culture blade. Morphology, location and presence or absences of internal organs were recorded.

A Distance between setae of a worm was measured by an ocular micrometer. Measurement of pineal or copulatory setae was also taken by an ocular micrometer under high magnification. Slides were prepared for studying spermathecae and different kinds of setae. All illustrations were drawn with the help of a camera lucida.

Culturing of earthworms

Among the collected earthworms some of them were segregated to culture under favourable laboratory condition. They were cultured in suitable cement tanks (1m length × 1 meter breadth × 50cm height) by providing the feeding material consisting of 25% decomposed organic waste and 25% predecomposed cow dung and 50% garden soil. A pool of 20 healthy matured worms was selected to extract the coelomic fluid for the study of antibacterial activity.

Cleaning and removing of gut content of earthworms

Fully matured earthworms were washed in running tap water in order to remove the sand particles from the surface of earthworms. Then the washed earthworms were soaked in N-saline solution and the solution was exchanged every time after wash till the gut of earthworms was thoroughly cleaned (Abhishek mathur et al., 2010).

Extraction of coelomic fluid

Coelomic fluid was extracted by applying a potential difference across a pool of worms, causing them to extrude coelomic fluid. A pool of twenty worms were placed in a large petridish in an electrolyte solution (0.1% NaCl) that was also served to collect the extruded coelomic fluid (ECF), and a potential difference of 9 V was applied across the worms for 20[^]30 s. The coelomic fluid was collected with the help of micropipette. The collected coelomic fluid was centrifuged for 5 minutes at 350 rpm and the filtrate was used. The fluid was used at the concentration of 100% to study the antibacterial activity (Bundy et al., 2001).

Antibacterial assay

Disk method was used to assay the antibacterial activity of coelomic fluid of *Eudrilus eugeniae*. Bacterial cells were grown overnight in LB medium and inoculated into 5ml of molten 0.6 g/l agar with a final concentration of 10⁶ colony forming units per ml, which was overlaid onto a 90mm petridish containing 10ml of 2g/l LB solidified agar. After the top agar hardened, sterilized blotting paper (about 6.0 mm in diameter), free from antibacterial activity, were impregnated with 20 μ L of coelomic fluid to be tested and placed on agar dishes inoculated with one bacterial strain. The dishes were incubated over night at 37.0°C. Control tests were performed with penicilin.

Bacterial strains used for determining antibacterial activity included *Escherichia coli*, *Staphylococcus aureus* and *Pseudomonas aeruginosa* which were obtained from Clab cultural laboratory, Trichirappalli, Tamilnadu, India. Antibacterial activity was determined by determining the Relative Magnitude of Inhibition (RMI). The data were statistically analyzed at 5.0% level employing ANOVA.

$$RMI = \frac{\text{Area defined by zone of inhibition (including disc)}}{\text{Area defined by the disc}}$$

Results and Discussion

Table 1 shows the antibacterial activity of the coelomic fluid of the epigeic species of earthworm *Eudrilus eugeniae* and the anecic species of earthworm *Polypheretima elongata* against two strains of gram positive bacteria namely *Staphylococcus aureus* and *Pseudomonas aeruginosa* and one strain of gram negative bacterium namely *E. coli*. Among the selected bacterial strains the maximum inhibition zone of coelomic fluid of *Eudrilus eugeniae* (7.01 ± 0.28mm) and *Polypheretima elongata* (2.43 ± 0.80mm) was noticed against the same species of bacterium *Staphylococcus aureus* when compared with the inhibition zones produced by other bacterial strains tested in the present study. The maximum antibacterial activity in the coelomic fluid may be due to the presence of the range of humoral defence factor including agglutinins (e.g. lectin), lysosomal enzymes (e.g. acid phosphatase) and bactericidins (Cooper, 2005). But at the same time the coelomic fluid of *Polypheretima elongata* exhibited the minimum inhibition zones (2.43 ± 0.80mm, 1.35 ± 0.19mm and 2.28 ± 0.29mm) against all the bacterial strains tested than the inhibition zone produced by the coelomic fluid of *Eudrilus eugeniae* against these strains (Plate 1, 2,3,4,5 and 6).

Besides, the antibacterial activity in the coelomic fluid of both species of earthworms against *Pseudomonas aeruginosa* did not make much difference in the size of zone of inhibition. As for *E.coli* *Eudrilus eugeniae*'s coelomic fluid showed higher inhibition zone against this bacterium than the zone produced by the coelomic fluid of *Polypheretima elongata*. According to control experiment the maximum and minimum inhibition zones were noticed against *Pseudomonas aeruginosa* and *Escherichia coli* respectively. However, the coelomic fluid of *Eudrilus eugeniae* showed high inhibition zone against *Staphylococcus aureus* and *Escherichia coli* than the control experiment whereas the control test revealed high inhibition activity against *Pseudomonas aeruginosa* than the coelomic fluid of *Eudrilus eugeniae*. But at the same time the control experiment revealed high inhibition zone against all the bacterial strains than inhibition zone of coelomic fluid of *Polypheretima elongata*. Dhanam et al. (2016) also suggested that the antimicrobial function of earthworm skin extraction to *S. marcescens* exposure was highest and the diameter of the growth of inhibition was 3mm whereas *S. aureus*, *K. pneumonia*, *B. aryabhatai*, *B. megaterium*, *P. putida* did not have the activity. Moreover, in the present investigation it has been proved that on the whole *Staphylococcus aureus*'s activity is highly inhibited by the coelomic fluid of both species of earthworms than the other bacterial strains tested.

The earthworm is a first terrestrial invaded organism and it is living in the soil with millions of microorganisms. During the microbial infection the immune system in the earthworms has evolved to protect them from pathogens, microbes, viruses and infected cell. Popovic et. al. (1996) have reported that the antibacterial activity in the coelomic fluid of earthworm may be due to the presence of innate immune mechanism and detect the microorganisms by recognizing conserved molecular pattern. The present study corroborate with those of Ramasamy et al.(2008) and Zasloff (2002), who stated that anti-microbial peptides constitute a

very important component of the innate immune system of *Eudrilus eugeniae* and *E.foetida* respectively.

Invertebrates in general and earthworm in particular apparently lack lymphoid organs, lymphocytes and immunoglobulins but they have very efficient host defence system, because the body fluids contain a range of humoral defence factors including agglutinins (e.g. lectin), lysosomal enzymes (e.g. acid phosphatase) and bactericidins (Cooper, 2005). Earthworms have primitive non-specific recognition systems, and they are mediating through innate immune responses. Earthworms have extremely effective physiochemical barriers and are acting as first line of defence against pathogens. During infection, group of leucocytes, free within blood vessels, or occupying fluid filled body cavities known as coelom, are ejected out through dorsal pores, bind to microbes, engulf and then kill them. The coelomocytes of earthworm otherwise known as leucocytes (monocytes, macrophages polymorphonuclear and neutrophils) are responsible for innate immune functions such as phagocytosis and encapsulation against pathogens and parasites. With this effect the worms are able to survey the danger exposures necessitated by their more sluggish habit (Renganathan.2006).

Earthworms kill microorganism by recognizing conserved molecular patterns (lipopolysaccharides (LPS) or peptidoglycans from bacterial cell walls, β -1, 3-glucan of fungal cell walls, and double stranded RNA of viruses) on pathogen's body surface. Recognition of molecules for foreign material has been named as pattern-recognition proteins (PRPs) (Medzhitov and Janeway, 1997) because the host's primitive effector cells would recognize molecular patterns rather than particular structures of the invading microorganisms. When bacteria invade into the coelomic cavity of earthworm, the coelomocytes starts to connect with each other by their adhesive structures around the bacteria and form "brown bodies" (Valembios, et al., 1982). At the same time the coelomocytes intensively synthesize and secrete proteins that adhere to the bacteria, forming aggregations and may inhibit their further proliferation. These proteins attach to the lectin like monosaccharide of the cellular membrane of the bacteria. Different proteins and peptides of different species of earthworms have been extensively studied and different mechanisms of actions have been proposed (Marta et al., 2010). Hence, it is very difficult to define which molecule and mechanism of coelomic fluid or extract of earthworms is responsible for its antibacterial activity.

Milochau et al (1997) revealed the anti-bacterial activity of dialyzed coelomic fluid of *Lumbricid*, *Eisenia foetida* Anderi. The antibacterial activity of coelomic fluid of *Eisenia foetida* was proposed by Valem et al (1982). Liu et al., (2004) identified and purified a novel antibacterial peptide namely OE3121 which is responsible for the efficient immune response of earthworm. It has been reported that the coelomic cytofactor I isolated from the coelomic fluid is involved in the activation at prophenoloxidase cascade via recognition at gram negative bacterial cell wall molecules, such as glucan and lipopolysaccharide (Hrzenjak et al. 1992). The present investigation states that it is difficult to define which molecules of coelomic fluid are responsible for its antibacterial activity. Hence, the further detail study is needed for screening out the exact molecules which are responsible for its antibacterial activity.

Plate 1. Inhibition zone of coelomic fluid of earthworm, *Eudrilus eugeniae* against *Staphylococcus aureus*

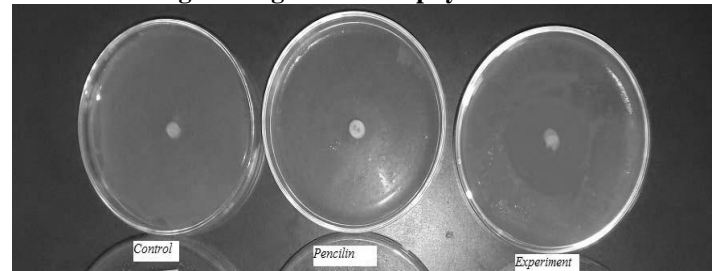


Plate 2. Inhibition zone of coelomic fluid of earthworm, *Eudrilus eugeniae* against *Pseudomonas aeruginosa*

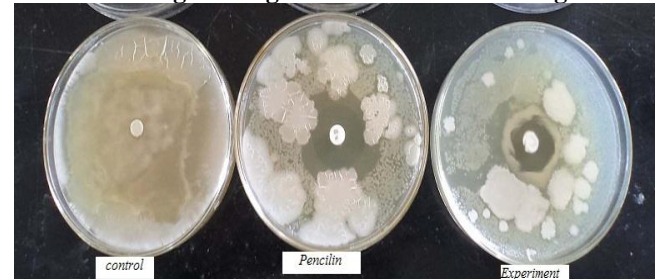


Plate 3. Inhibition zone of coelomic fluid of earthworm, *Eudrilus eugeniae* against *E.coli*.

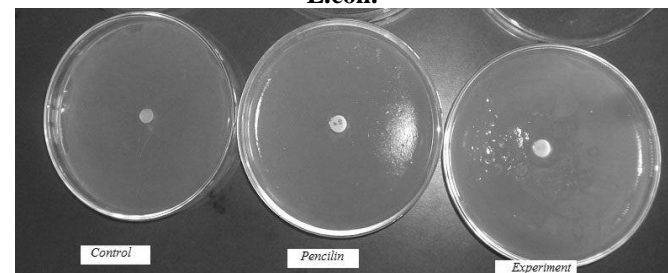


Plate 4. Inhibition zone of coelomic fluid of earthworm, *Polypheretima elongata* against *Staphylococcus aureus*

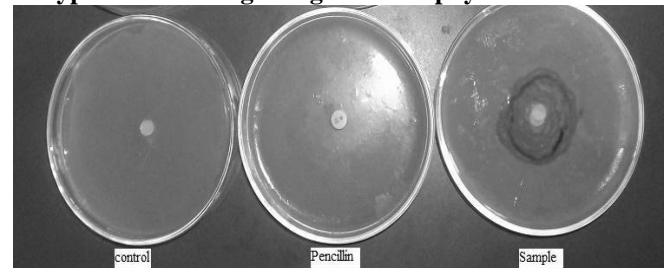


Plate 5. Inhibition zone of coelomic fluid of earthworm, *Polypheretima elongata* against *Pseudomonas aeruginosa*

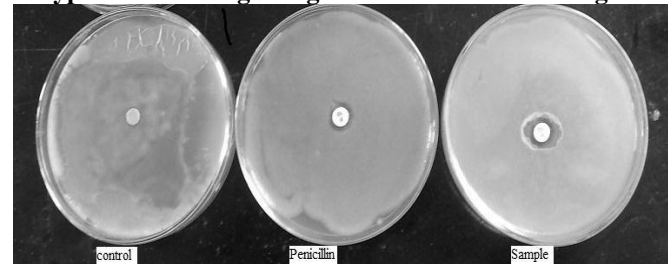


Plate 6. Inhibition zone of coelomic fluid of earthworm, *Polypheretima elongata* against *E.coli*.

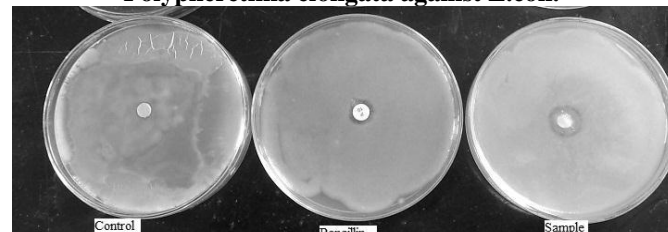


Table 1. Comparative study on antibacterial activity in the coelomic fluid of *Eudrilus eugeniae* and *Polypheretima elongata* against different species of bacteria. Size (mm) of the zone of inhibition produced by 6mm disc on bacterial lawn.

Species of bacteria	Inhibition zone (mm) of coelomic fluid of <i>Eudrilus eugeniae</i>			Inhibition zone (mm) of coelomic fluid of <i>Polypheretima elongata</i>		
	A1	A2	A1/A2=RMI	A1	A2	A1/A2=RMI
Control	0	0	0	0	0	0
I Gram- positive - <i>Staphylococcus aureus</i>	42.60	6.0	7.01	14.60	6.0	2.43
- <i>Pseudomonas aureus</i>	14.10	6.0	2.35	13.60	6.0	2.28
II Gram-negative - <i>E.coli</i>	16.0	6.0	2.66	8.10	6.0	1.35
III Antibiotic (Penicillin)	Zone of inhibition(mm) of antibiotic(Penicillin)					
	A1		A2	A1/A2 = RMI		
- <i>Staphylococcus aureus</i>	21.0		6.0	3.70		
- <i>Pseudomonas aureus</i>	21.4		6.0	3.97		
- <i>E.coli</i>	6.6		6.0	1.77		

A1- Area defined by zone of inhibition (including disc) A2- Area defined by the disc RMI- Relative magnitude of inhibition

Statistical analysis for table 1

S.No	Species of bacteria	Zone of inhibition(mm) of coelomic fluid of <i>Eudrilus eugeniae</i>	Zone of inhibition(mm) of coelomic fluid of <i>Polypheretima elongata</i>
1.	I- Gram positive - <i>Staphylococcus aureus</i> - <i>Pseudomonas aureus</i>	7.01 ± 0.28 2.35 ± 0.31	2.43 ± 0.38 2.28 ± 0.29
2.	II- Gram negative - <i>E.coli</i>	3.66 ± 0.31	1.35 ± 0.19
3.	III- Antibiotic (Penicillin) - <i>Staphylococcus aureus</i> - <i>Pseudomonas aureus</i> - <i>E.coli</i>		3.70 ± 0.13 3.97 ± 0.24 1.77 ± 0.44

Values are explained by mean ± SD of six samples - significant at 5% level

References

Balamurugan, M., K. Parthasarathi, E.L. Cooper and Ranganathan, L.S. 2008. Anti-inflammatory and anti-pyretic activities of biologically active extract isolated from whole earthworm, *Lampito mauritii*. J. Ethnopharmacol. 121(2):330-2.

Bristow, H.S. 1932. Insects and other invertebrates for human consumption in Siam. Trans Entomol soc (London), 80: 387-404

Bruhn H., Winkelmann J., Andersen C., Andra J. and Leippe M. (2006) Dissection of the mechanisms of cytolytic and antibacterial activity of lysenin, a defence protein of the annelid *Eisenia fetida*. Developmental and Comparative Immunology, 30: 597-606.

Bundy, J.G., Osborn, D., Weeks, J.M., Lindon, J.C., Nicholson, J.K., (2001). An NMR-based metabonomic approach to the investigation of coelomic fluid Biochemistry in earthworms under toxic stress. FEBS letters 500, 31-35.

Carr LGK. Interesting animals, foods, medicines and omens of the eastern Indian with comparison to ancient Europe practice. Journal Washington Academy of Science. 1951; 41,229-235.

Cho JH, Park CB, Yoon YG, Kim SC. Lumbricin I, Biochim. Biophys. Acta, 1998; 1408: 67-76.

Cooper EL, 2005. CAM, eCAM, bioprospecting: the 21st century pyramid. Evidence Based Complementary Ancient Medicines 2:125-127.

Cooper, E.L., Kauschke and E., Cossarizza., 2002. Digging for innate immunity since Darwin and Metchnikoff. Bioassays, 24: 319-333.

Çotuk A. and Dales P.R. (1984) The effect of the coelomic fluid of the earthworm *Eisenia foetida* sav. on certain bacteria and the role of the coelomocytes in internal defense. Comparative Biochemical Physiology, 78A (2): 271-275.

Dhanam S, Arumugam T, Rameshkumar N, Krishnan M, Kayalvizhi N (2016) Antimicrobial Potential of Earthworm *Wegeneriona* sp. against Human Pathogens. J Anal Pharm Res 3(4): 00060. DOI: 10.15406/japlr.2016.03.00060

Davie, E. W., Fujikawa, K. and Kisiel, W. (1991) The coagulation cascade: Initiation, maintenance, and regulation. Biochemistry 30, 10363-10370.

E. L. Cooper, and P. Roch, 2003. "Earthworm immunity: a model of immune competence," Pedobiologia., vol. 47, pp. 676-688.

Edwards, C. A. and Lofty, J. R. 1972. Biology of earthworms. Chapman & Hall, London. 283 pp.

Edwards, C. A., Bohlen, P. J., Linden, D. R. and Subler, S. 1995. Earthworms in agroecosystems. In: Earthworm Ecology and Biogeography in North America. (Hendrix, P. F. eds.), Lewis Publisher, Boca Raton, FL, pp: 185-213.

Edwards, C.A and P.J. Bohlen. (1996) Biology and Ecology of Earthworms, 3rd ed, Chapman and Hall, London.

Guerrero, R.D. 1983. The culture and use of *Perionyx excavatus* as a protein resource in the Philippines pp. 309-319. In: J.E. Satchell (ed.) Earthworm Ecology, Chapman and Hall, London. 495 p.

Harwood, M. and Sabine, J.R. 1978. The nutritive value of worm meal. Proc 2nd Austr poul stockfeed conv, Sydney pp:164-171.

Harwood, M. 1976. Recovery of protein from poultry waste by earthworm. Proc 1st Austr poul stockfeed conv, Melbourne. pp: 138-143

Hong, C., 2007. Earthworm fibrinolytic enzyme and anti-tumor activity on human hepatoma cells, in vitro and in vivo. Chinese Med. J. 260-264.

- Hrzenjak, T., M. Hrzenjak, V. Kasuba, P. Efenbermarnculic, s. Levanat 1992. A new source of biologically active compounds-earthworm tissue (*Eisenia foetida*, *Lumbricus rubellus*). *Comp. Biochem. Physiol.* 102: 441-447.
- Julka, J.M. and Paliwal, R. (1993) A new species of Perionyx Perrier (Megascolecidae, Oligochaeta) from northwest Himalaya. *India. J. Bombay nat. Hist.Soc.*, 90:461-462
- Julka, J.M.1988. The fauna of india and adjacent countries (Haplotaxida:Lumbricina;Megascolecoidae:Octochaetidae). Zoological survey of India, culcutta.400pp
- Kathireswari, P.,Alakesan,A.,Abirami,P., and Sangeetha,P. 2014. Antimicrobial activity of Earthworm Coelomic fluid against disease causing microorganism. *Int.J.Curr.Microbial.App.Sci* (2014) 3(8): 608-613.
- Kim YS, Kim YE, Byun HS and Chang CS. Regulation of NAD⁺ glycohydrolase activity by ADP ribosylation. *J. Biochem. Mol. Bio.*, 1995; 28, 398.
- Kohlerova, P., Beschin, A., Silerova, M., De Baetselier, P. and Bilej, M., 2004. Effect of experimentall microbial challenge on the expression of defense molecules in *Eisenia foetida* earthworm. *Developmental and comparative Immunology*, 28:701-711.
- Lange S, Kauschke E, Mohrig W, Cooper EL. Biochemical characteristics of Eiseniapore, a pore-forming protein in the coelomic fluid of earthworms. *Eur J Biochem* 1999;262:547-56.
- Lange S., Nubler F., Kauschke E., Lutsch G., Cooper E.L. and Herrmann A. (1997) Interaction of earthworm hemolysin with lipid membranes requires sphingolipids. *The Journal of Biological Chemistry*, 272(33): 20884-20892.
- Lassegues M., Roch P., Valembois P and Davant N. (1981) Patogenic action of some bacterial strains in the lumbricia *Eisenia fetida Andrei*. *C.R. Académie des Sciences Paris D.*, 292: 731-734.
- Leipner C, Tuckova I, Rejnek J and Lagner J. Serine proteases in coelomic fluid of annelids *Eisenia fetida* and *Lumbricus terrestris*. *Comp. Biochem. Physiol.* 1993; 105 B, 679.
- Macdonald, D.W.1983. Prediction on earthworms by terrestrial vertebrates, pp.393-414, In: J.E. Satchell (ed.) *Earthworm Ecology*, Chapman and Hall, London.495 p.
- Marta J, Fiołka Mirosław P, Zagaja Tomasz D, Piersiak Marek Wróbel and Jarosław Pawelec. *Journal of Invertebrate Pathology*, 2010, 105 , 63-73.-254.
- Medzhitov R, Preston-Hurlburt P, Janeway CA Jr. *Nature*. 1997; 388 (6640):394-7.
- Mekada, H.N. Hayashi, H. Yokota and J.Okumura. 1979. Performance of growing and laying chickens fed diets containing earthworms (*Eisenia foetida*). *Jpn. Poult. Sci.*16:293-297.
- Mihara, H., R. Ikeda and Yonnet, T. 1996. The Useful of Earthworm Powder. Miyazaki Medical College, Kiyotake, Miyazaki.
- Milochau A, Lassègues M and Valembois P 1997 Purification, characterization and activities of two hemolytic and antibacterial proteins from coelomic fluid of the annelid *Eisenia foetida Andrei*; *Biochim. Biophys. Acta* 1337 123–132
- Nagasawa H, Sawaki K, Fuji Y, Kobayashi M, Segawa T, Suzuki R, Inatomi H. *Anticancer Res.* 1992: 1061
- Nagasawa H, Sawaki K, Fuji Y, Kobayashi M, Segawa T, Suzuki R, Inatomi H, 1991. Biology of lysenin a protein in the coelomic fluid of earthworms. *Anticancer Res.* 1061.
- Noda N, Tsunefuka S, Tanaka R, Miyahara K, 1992. Effect of an earthworm, *Lumbricus rubellus*. *Chem. Pharm. Bull.*; 40:2756.
- Ogata A and Mori HJ. 1938. Constituents of the earthworm as an antipyretic agent I. *Journal of Pharmacological society of Japan.* (58): 859-87.
- Ogata A, Morimoto K and Mori H J. Constituents of the earthworm as an antipyretic agent II. *Ibrid.* 1939; 59, 481-494.
- Ogata, T. 1938. Note preliminaire sur deux especes nouvelles de trematodes du genre *Astiotrema*, provenant de l' *Amyda maackii*. *Zool. Mag. Japan.*, 50(1): 50-52.
- Oleynik AS and Byzov BA. Response of bacteria to earthworm surface excreta. *Microbiologiya.* 2008; 77, 854-862.
- Pan W., Liu S .,Ge F. and Zheng T. 2003. Reconfirmation of antimicrobial activity in the coelomic fluid of the earthworm *Eisenia foetida Andrei* by colorimetric assay, *J. Biosci.*, 28 (6): 723-731.
- Popovic, M., Lj. Tiska-Rudman, T. Hrzenjak 1996. Tissue extracts of earthworm *Eisenia foetida* (G-90) as a blood anticoagulant and fibrinolytic. *Vet. Arhiv.* 66: 161-167.
- Prabha, M.L., I.A. Jayaraaj, R. 2007. Jeyaraaj and S. Rao: Comparativestudies on the digestive enzymes in the gut of earthworms, *Eudrilus eugeniae* and *Eisenia fetida*. *Indian J. Biotechnol.*, 6,567-569.
- Ramasamy, P.K.,Jeyaraj,R.,Indira A.J., and Sridevi, S. 2008. Antimicrobial activity in the Coelomic fluid of the earthworm *Eudrilus eugeniae*. *Asian Jr. of microbial. Biotech.sc.* 10(4): 927-929.
- Ranganathan, L.S. 2006. Vermicomposting enhances huminification, mineralization and chelation. *J. Ann. Univ. Science*, 42:1-14
- Sabine, J.R. 1978. The nutritive value of earthworm meal. In: *Proceeding of the Conference on Utilization of Soil Organisms in Sludge Management*. Syracuse N.Y.pp. 122-130. Edited by R.Hartenstern. S.U.N.Y., Syracuse, U.S.A.
- Senapati, B.K. 1992. Vermitechnology. An option for recycling of cellulosic waste in India. In: *New trends in biotechnology.* (Eds. Subba Rao N.S., Balagopalan C. and Ramakrishnan S.V). pp 347-358. Oxford and IBH Publishing, New Delhi, India.
- Shobha, S.V., and Kale, R. 2007. Antimicrobial potency of earthworm, *Eudrilus eugeniae* on certain plant pathogens. *Administrator* .
- Stafford, E.A. and A.G.J Tacon. 1988. The use of earthworms as a food for rainbow trout, *Salmo gairdneri*, pp. 193-200, In: C.A. Edwards and E.F. Neuhauser (eds.) *Earthworms in Waste and Environmental Management*, SPB Academic Publishing, The Hague, The Netherlands.
- Stephenson, J.1930. *The Oligochaetidae*. Oxford university press, London. 1977pp.
- Tang, Y. Zhang, J Gui, L. et al 2000. Crystalization and preliminary X-ray analysis of earthworm fibrinolytic enzyme component A from *Eisenia foetida*, "Actacrytallographica D, Vol. 56, no. 12 pp. 1659-1661.
- Valembois P, Roch P, Lassègues M and Cassand P. 1982. Antibacterial activity of the hemolytic system from the earthworm *Eisenia foetida Andrei*; *J. Invertebr. Pathol.* 40 21–27
- Vohora, S.B. and Khan, M.S.Y. 1978. Animal origin drugs used in Unani medicine. *Institute of History of Medicine and Medical Research*, Tughlaqabad, New Delhi. 137pp.

Wang JD, Narui T, Kurata H, Takouchi K, Hashimoto T, Okuyama T, 1989. Fibrinolytic activity of the earthworm extract. *Chem. Pharm. Bull.*; 37: 2236.

Woo, J.-I., Bahk, Y.-K., Yu, S.-R., Paik, S.-R. and Chang, C.-S. 1996. Evidence for existence of a water-extractable anticoagulant in an earthworm, *Lumbricus rubellus*. *J. Biochem. Mol. Biol.* 29, 500-506.

Xu, J.U. et al. 1984. Substitution of fishmeal with earthworms in broiler diet. *Chinese J. of Ani. Sci.*: 1,9-11.

Yegnanarayan, R., P.P. Sethi, P.A. Rajhan, K. Pulandiran and Ismail, S.A. 1987. Anti-inflammatory activity of total earthworm extracts in rats. *Indian J. Pharmacol.* 19:221-224.

Zasloff, M. 2002. Antimicrobial peptides of multicellular organisms. *Nature.* 389-395.