



Leaf Spot on Hybrid Varieties of Asteraceae Causes 12% loss in Productivity in Nepal & India

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ABSTRACT

Dwarf varieties of *Helianthus annuus*, *Tagetes erecta*, *T. Patula* and other plants belonging to Asteraceae are cultivated as ornamental garden plants in Nepal and India. Leaf spots and blight, downy mildew, mycoflora of floral parts, collar rot, Necrosis, charcoal rot and other of Asteraceae already reported. Leaf spots caused by the pathogen *Albugo spp.* bring 12% loss in productivity. Akoijam & chandel (2010), Bhatia et.al (2005), Biga(1955), chander (2003), Dube (1985), chavhan et.al (2008), kumar & Dewivedi (1981), Kulkarni et.al (2007), Lakhmod et.al (2007&2007), Mayee & Datur (1986), Pandey & Tewari (2017), Singh et.al (1999), Kulkarni et.al(2007), Singh et.al (1999), Singh (1980) , Sinha (2011), Suryawanshi et.al (2015) & Thirumalachar et.al (1949) have well studied Asteraceae and possible pathogens in their own ways.

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1. Introduction

Dwarf Varieties of *Helianthus annuus*, *Tagetes erecta* & *T. patula* belonging to Asteraceae are cultivated as ornamental garden plants in Nepal & India. Such Plants are available in May month in Nursery of Biratnagar, where imported from Silligauri or kolkata (west Bengal) India. White leaf spots, showing plants of Asteraceae selected for study. 6 plants at Jogbani (India) on Date 26/05/2019 and 5 plant at Biratnagar (Nepal) Dwarf varieties of *H.annuus* – red, pink,brown thin ray floret and brown thick ray floret, all had white leaf spots, while *T. erecta* & *T. patula* showed later the presence of leaf spot on date 28/06/2019 were Planted in pots and garden respectively.

2. Review of Literature

Akoijam & chandel (2010)¹ reported resistance against leaf spot and flower blight caused by *Alternaria zenniae* Pape, while Bhatia et.al (2005)² mentioned enhancement of plant growth and suppression of collar rot of sun flower caused by *Sclerotium rolfsii* through fluorescent *Pseudomonas*. Biga (1995)³ described key- identification of *Albugo spp.* chander Rao (2003)⁴ mentioned hybrid seed – production technology in sunflower. Dube (1985)⁵ wrote the book ' An introduction to Fungi ' following Ainsworth-classification. chavhan et.al (2008)⁶ reported association of new fungal species with leaf spot and blight of Sunflower. Kumar & Dwivedi (1981)⁷ mentioned mycoflora association with floral parts of sunflower. Kulkarni et.al (2007)⁸ described molecular variability in *Plasmopara halstedii* causing downy mildew of sunflower. Lakhmod et.al (2007)⁹ reported sunflower Necrosis - Transmission & host range studies & again Lakhmod et.al (2007)¹⁰ reported epidemiology of sunflower Necrosis disease. Mayee & Datur (1986)¹¹ mentioned host and disease assessment scale-sunflower in Phytopathometry. Pandey & Tewari (2017)¹² described selection of host differentials for the identification of *Albugo candida* phenotypes. Singh et.al (1999)¹³ reported a stain of cucumber mosaic cucumo-virus, causing mosaic in marigold in India. Singh (1980)¹⁴ wrote the book

'Introduction to principles of plant pathology' related the relationship of pathogen and host in different conditions. Sinha(2001)¹⁵ described fungi- associated with infected pollen gains Of marigold (*T. erecta*) suryawanshi (2015)¹⁶ mentioned , managing *Macrophomina Phaseolina* causing charcoal rot of sunflower (*Helianthus annuus*) by soil-amendment. Thirumalachar et.al (1949)¹⁷ reported gametogenesis and oospore formation in *cystopus* (*Albugo*) *evolvuli*.

3. Morphological symptoms

Plants of Hybrid – Asteraceae potted in eastern direction (p1 p2p3&p4)) While 2 plants (P5&P6) in western direction at Jogbani, Bihar, India on Date 26/05/2019 (P7P8P10P11) in eastern direction while P9 – one plant in western direction were grown at Biratnagar, province NO.1, Nepal. All plants showed the white leaf spots being several in no. were present on Date: 30/05/2019

Leaf spots-(+) On dorsal & ventral surfaces, elongated well developed on margin also, white transparent mycelia visible under sun light, shape of leaf spot is oval, circular, elongated, rectangular surrounded by dead tissues.

Further detail of study was done on Date 05/06/2019 at 8.45-10.20 A.M

Size of leaves-36cm²

lost area of productivity due to pathogens – 4.53 cm²

i.e loss of productivity = 12.6% Maximum leaves in P₆ -90 leaves

i.e loss of Productivity = 11.34% =12%

i.e Forecasting-12% Loss in productivity due to leaf spots.

Again variations of symptoms appeared on plants & their parts was studied on Date.13-06-2019 at 10.05-11.0 A.M given in the following table 2

Leaves- shrinkage, dried, white powdery mass leaf spots.

Stem- towards apex was yellowish, hollow-ridged, shrinkage & dried in P₁&P₂ but P₃- green in colour, in P₄ especially presence of white dots(Several in no.) aggregated giving cottony appearance.

Table No.1

I N D I A	E A S T E R N	Plants	Height	Nature	Leaf spots	Bud	flower	Remark
		P ₁	50cm.	Dry	(+)	(-)	dry 1(+)	
P ₂	54cm.	Dry	(+)	(-)	do			
P ₃	54cm.	Green	(+)	(-)	dry 2(+)			
P ₄	56cm.	Green	(+)	(+)	green 1(+)			
W E S T E R N	P ₅	67.5cm.	Dry green	(+)	Green 2(+)	dry2 (+)		
	P ₆	52.5cm.	green	(+)	Green 1(+)	1(+)		
NEPAL	EASTERN	P ₇	69cm.	Dry	(+)	(+)	dry 1(+)	
		P ₈	78.7cm.	Dry	(+)	(-)	dry 1(+)	
		P ₁₀	68.5cm.	Green	(-)	(-)	(-)	
		P ₁₁	36cm.	Green	(-)	(-)	(-)	
	Western	P ₉	39cm.	Green	(+)	(+)	(-)	
(+) presence & (-) absent								

Table No.2

E A S T E R N	PLANT	Height	Stem above soil showing symptoms
	P ₁	49.6 cm.	3cm. broken black & 11 cm White Powdry mass.
P ₂	49.2 cm.	4.2cm. broken, 4.2cm. white powdry mass & 6 cm. black .	
P ₃	52.0 cm.	8cm black & a little white powdry mass	
		5.5cm. black& white	
WESTERN	P ₅	64.0 cm.	5.5cm. black & white
	P₆	63.0 cm.	14cm. black & Green

Leaves in P₄ were same (as in P₁, P₂,P₃&P₅) but 3

Leaves were very minute-

1 largest – 1.3 cm. white & enrolled.

1 Intermediate – 0.8 cm white.

1 smaller – 0.3 white.

These leaves were collected to microscopic study.

In P₆ plant, leaves- green & with white spots.

Date: 17-06-2019 at 7.42-8.03AM

i. Symptoms showing a flower was broken from P6 Plant & 12 ray florets were studied.

ii. Length – 1.8 cm (lower portion was 0.8cm)

iii. White leaf spots- elongated, oval a dots-(+)

iv. Dorsal side of ray florets- 8 in no, contain raised, oval or deposition of tissues – (+).

v. Central side in all 12 ray florets showed white raised tissue – (+)

vi. Lower portion of central side showed white mycelia – (+)

Date. 17/06/2019 at 4.45-6.21 P.M

I. Dry root- length 11.3cm, elongated taproot- 9.5 cm, sec. root-2.9, tertiary- 1.2cm. Total no. of roots-32 i.e.32+1=53

II. Colour – white, diameter – 0.5cm & area – 0.196cm²

III. Diameter of vascular region – 0.2cm& area – 0.785cm², in which 0.059cm² is black & 0.0196cm² is white.

Date: 02/07/2019 at 11.30-12.45 PM

Place–Dept.of Botany, M.M.A.M CAMPUS, BIRATNAGAR, T.U, NEPAL.

Microscopic observation –

a. Root- dry, hard, white hairs with tap root.

- taproot under t.s- vascular degeneration and filled with blackmass and white structure. i.e Presence of *Macrophomina* spp. , Suryawanshi et.al (2015)16 and some portion of soil.

b. Leaf spot- White, internal yellowish tissue (infected portion)

- Mycelia (+), conidia – numerous (+)

- oogonium (+)

c. Flower – Dry & white spot (+)

d. Ray-floret– numerous mycelia at periphery with white spot (+)

-Mycelia (+), Conidia (+) & chamydospores (+).

e. Disc-floret – White spot (+) bunch of mycelia (+), Conidia (+).

f. Leaves of P7&P8- White spot (+), very minute but visible.

- Conidia (+), Germinating Conidium (+)

- Aggregation of several conidia in a membranous ball attached with mycelium (+).

4. Conclusion

The dwarf variety of *Helianthus annuus* comprise white leaf spot, which results 12% loss in Productivity due to *Albugo spp.* and other fungi especially responsible to vascular disintegration and becomes causes of dryness or death of the plant.

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References

1. Akoijam, R.S & chaudel, s (2010) screening of some marigold cultivars (*Tagetes erecta* & *T.patula*) for resistance against leaf spot and flower blight caused by *Alternaria zinnia* Pape. *Indian Phytopath.* V63(3) IPS, IARI, New Delhi-110012, p354-355.
2. Bhatia, S et.al (2005) Enhancement of plant growth and suppression of collar rot of sunflower caused by *Sclerotium rolfsii* through fluorescent *Pseudomonas*, *Indian Phytopath.* V58(1) P.17-24.IPS, IARI, New Delhi-110012, India.
3. Biga, M.L.B (1955) Riesaminazione delle specie del genere *Albugo* in base alla morfologia dei conidi *sydowia* 9 p.358-399.
4. Chander Rao, s (2003) Hybrid seed production technology in sunflower, In sunflower in India, Directorate of oil seeds Research, Hyderabad, Indian Council of Agricultural Research, India p-112.
5. Dube, H.C (1985) An Introduction to fungi vikash publishing House Pvt .Ltd, 5- Ansari Road, New Delhi-110002.
6. Chavhan, R.L et.al (2008) Association of new fungal species with leaf spot and blight of sun flower and cloning of their ribosomal RNA genes. *Indian Phytopath.* V61(1) p.70-74. IPS, IARI, New Delhi-110012, India.
7. Kumar, V & Dwivedi, R.S (1981) Mycoflora associated with floral parts of sunflower. *Indian Phytopath.* V.34-317. IPS, IARI, NEW Delhi-110012, India.
8. Kulkarni, s et.al (2007) Molecular variability in *Plasmopara halstedii* causing downy mildew of sunflower. *Indian phytopath.* V60(1) p.48-51. IPS, IARI, New Delhi -110012, India.
9. Lakhmod, L.k et.al (2007) sunflower Necrosis Disease. *J.Pl.Dis.Sci.* Vol2(1) p.85-87, Asso. of plant pathologist (Central India) Dr.Punjabrao Deshmukh krishi vidyapeeth, Akola-144104. India.
10. Lakhmod, Lk et.al (2007) Epidemiology of sunflower Necrosis Disease. *J.pl.Dis.* Vol 2 (1) p.115-116 Asso.of plant pathologist (central India) Dr.Punjab rao Deshmukh krishi vidyapeeth, Akola-444104, India.
11. Mayee, C.D & Dafar, V.V (1986) Host and disease assessment scales-sunflower. In phytopathometry, Technical Bulletin No.1, M.A.U, Parbhani, India P.93.
12. Pandey, P & Tewari, A.K (2017) selection of host differentials for the identification of *Albugo candida* phenotypes. *Indian Phytopathology* V70(3) p.375-380.IPS,IARI, New Delhi 110012 India.
13. Singh, D et.al (1999) A strain of Cucumber mosaic cucumovirus, causing mosaic in Marigold in Indian Phytopath. IPS, IARI, New Delhi -110012 p.114-117.
14. Singh, R.S (1980) Introduction to Principles of plant pathology. Oxford & IBH Publishing Co. 66 Janpath, New Delhi-110001, India.
15. Sinha, A.A (2011) fungi associated with infected pollen grains of marigold (*Tagetes erecta*). *Indian Phytopath.* V64(2) p.206.IPS, IARI, New Delhi-110012, India.
16. Suryawanshi, A.P et.al (2015) Managing *Macrophomina Phaseolina* causing charcoal rot of sunflower (*Helianthus annuus*) by soil amendment. *Indian Phytopath.* V.68(2) p.196-200.IPS, IARI, New Delhi-110012, India.
17. Thirumalachar, M.J et.al (1949) Gametogenesis and oospore formulation in *cystopus (Albugo) evolvuli*. *Bot.Gaz.* 110: P.487-491.