55040



K. K. Mishra/ Elixir Appl. Botany 149 (2020) 55040-55042 Available online at www.elixirpublishers.com (Elixir International Journal)





Elixir Appl. Botany 149 (2020) 55040-55042

Fusarium Oxysporum on Rosa Species in Eastern Zone of Nepal and North Bihar, India

K. K. Mishra

Department of Botany, M.M.A.M.Camus, Biratnagar, T. U., Nepal.

ARTICLE INFO

Article history: Received: 5 November 2020; Received in revised form: 1 December 2020; Accepted: 8 December 2020;

Keywords

Fusarium oxysporum Rosa sp,

ABSTRACT

9 potted plants of *Rosa sp.*, in which 3 *R. indica* L. and others hybrid *Rosa sp.* 10 & 2 years old respectively werye observed the sequence of formation of dried or dead stems and alive stems. New branches and bud formed after cutting on date 6/11/2020 and the observation on date 27/11/2020 i.e. within 21 days. *Fusarium oxysporum* causing the wilting or dried condition in *Rosa sp.* was confirmed in the Laboratory of Department of Botany, M.M.A.M. CAMPUS, BIRATNAGAR, T.U, U NEPAL on Date 22/11/2020 at 1-2.15 P.M as characterised by Synder & Hansen (1940, 1941,1945)^{27,28,29} Synder (1965)³⁰, Dube (1983)³ & Mehrotra (1980)¹¹.

© 2020 Elixir All rights reserved.

1. Introduction

Rosa indica L.is well known ornamental plant. Its petal is useful to get rose water, essential oil, gulkund & perfume. Several hybrid varieties are preferred in garden. Worship purposes at home or temple is also common practice and usual manner to gift any one, whom loves or affection or even regard as noticed. It has good economic importance in our lives. It belongs to the family Rosaceae. Stem is woody, solid, green, aerial, erect, branched and prickly (spines), While the leaf is compound, petiolate and adnate stipule. Leaflets present in no. 3-5 even sometimes more, opposite, ovate, dentate, acute, unicostate reticulate venation. Solitary, axillary a clusters of Inflorescence is seen. Flowers are bracteate [sometimes(-)], pedicillate, complete, hermaphrodite, actinomorphic, perigyrous, large, Scented, Variously coloured & thalmus is cup-shaped. Calyx-sepals 5, gamo sepalous, lanceolate, deutate, hairy and green in colour, corolla Petals 5 or indefinite, polypetalous, rosaceous, imbricate & each petal is ovate type, Various colour, showy and fragrant. Androccium many i.e. polyandrous, inserted on the edge of the disc, unequal, dithecous, basifixed & introse. Gynoecium - poly carpellary, apocarpous, semi-inferior, ovule one in each carpel, basal placentation, style Lateral, hairy, free, stigma terminal. Fruit - Staerio of achenes, enclosed by the succulent bright red calyx tube called hip. Seed - non edospermic Sharma (1989). Rosa damascena Mill., R.centifolia L. and R. moschata Mill. are the Source of flower-petals to get essential oil in perfumery kochhar (1998). According to him 100 gms of flowers yields 0.5gm oil (even sometime Less) i.e. 0.4 -0.5% oil is obtained by the distillation mechanism and becomes costly. The principal constituent of rose oil is phenylethyl alcohol (soluble in water) and other components are citronellol, geraniol and nerol. Rose oil is largely used in perfuming or scenting Soaps, Soft drinks and liquers. Rose water is used in eye lotions and eye washes. It is sprinkled on guests on ceremonial occasions. Dried petals in making cold drinks during Summer is used. The fruits (rose hips) are source of vitamin C.

2. Review of Literature

The characteristics of host plant Rosa indica L. was described by several taxonomists and the use described by Sharma (1989) and Kochhar (1998), while the details in wealth of India. The description of the pathogen Fusarium oxysporums was given by wollen weber & Reinking³² in Berlin, while Synder & Hansen (1940, 1941 & 1945)^{27,28,29} mentioned species concept of Fusarium with references and differences again Synder (1965)³⁰ gave the Current status of Taxonomy and its Perfect stage. Some basic textbooks by different authors Singh (1968 & 1980)^{25,26} Mehrotra (1980)¹ Ingold $(1971)^6$ & Dube $(1983)^3$ have been giving the knowledge of *Fusarium sp.* Mishra $(2008)^{12}$ presented the paper at vishakhaptnam in ISCA-Conference. Mäshra (2012a, 2012b)^{13,14} presented two papers at Banglore, participating Indian Phyto pathological Society, IIHR, Hissarghatta lake post. Devi (2010)¹, Dorjey (2017)², Dubey (2016)⁴, Dubey et.al $(2017)^5$, kar et.al. $(2016)^7$, kaur et.al $(2019)^8$, khan et.al $(2017)^9$, Kotari et. al. $(2018)^{10}$, Mishra & Dhar $(2010)^{15}$, Mukhopadhyay et.al. $(2017)^{16}$, Murumkar $(2010)^{17}$, Mushrif & Khulbe $(2010)^{18}$, Nazia & Sharda $(2017)^{19}$, Ramappa & Gowri $(2016)^{20}$, Ray & Sharma $(2010)^{21}$ Ray et.al $(2016)^{22}$, Saharan $(2010)^{23}$, Singh et al $(2010)^{24}$ & wavare $(2017)^{31}$ have given different aspects, control, genetic behavior etc. about F. oxysporum. The phenomenon of wilting diseases in different host plants has been attracted by different mycologists at present.

3. Observation

i. 3 plants of Rosa, *indica L*, having 10 years old and 6 plants of hybrid *Rosa sps.* (used to give different varieted colours like Pink, Deepred, yellow, white-red patches, mixed colour & white orange) having 2 years old were cut on Date 6/11/2020.

ii. Both dried & Living Stems were cut Mannure of dried Cow dung were poured on date 4/11/2020.

^{© 2020} Elixir All rights reserved

No. of	Age of plants in	Dead bran	Alive	Bud	Remarks
Obs.	years.	ches. No.	branches No.		
1	10	3	1	3	Main 1cm., Lateral 2 -3 cm each
2	10	1	6	8	Whorl of branching, 15cm dia. & 8 buds
3	10	21	4	2	Maximum dead branches
4	2	4	2		One 5 cm. & another 4 cm.
5	2	7	11	1	Maximum new branches of different
					length
6	2	10	6		3-6 cm.
7	2	5	3	1	$1^{\text{st}} 30 \text{cm}, 2^{\text{nd}} \& 3^{\text{rd}} - 5 \text{cm},$
8	2	5	3		3cm, 5cm – 2
9	2	9	5		10cm – 18cm.

iii. The growth on living or alive stems were noted on Date 27/11/2020, which is given above in Table No. 1.

iii. The dried or dead branches are due to *Fusarium* oxysporum (var. rosae).

iv. Maximum No. of dead branches is 21 i.e. Maximum loss of stem in *R. indica L*.

4. Microscopic observation

Infected leaves and stems were Selected on Date 22/11/2020 at 1-2.15 P.M.

i. At - Laboratory of the Dept of Botany, M.M.A.M. Campus, Biratnagar, T.U., Nepal.

- ii. Leaf Mycelium (+).
- Micro-conidia in groups (+).
- colour slightly blackish at the base of leaflet (+)

- Accumulation of host cells with mycelia.

iii. Stem - Mycelium (+) in the vascular region i.e. in Xylem and phloem.

- also in cortex.

- Macroconidium is slightly curved & 4 segmented

- one segment, out of four is thick walled larger i.e. protesting unfavourable condition for germination i.e. chlamydospore

The identification was to be *Fusarimu oxysporum* var. *rosae* according synder Sander $(1965)^{30}$, Drube $(1983)^3$ & Mehrotra $(1980)^{11}$. The nine species concept of Synder $(1865)^{30}$ was accepted by mycologists.

5. Climatic condition

There is specification of the host *Rosa indica L*. and the pathogen *Fusarium oxysporum*. The maximum temperature and the minimum temperature is 36° C & 8° C respectively from Sept. 2019 to oct. 2020 and favours the disease wilting in the host.

6. Conclusion

Maximum loss in plant due to dried branches, gives less productivity and artificially, cut in *Rosa indica*, Blocking due to deposition of mycelia, macrocoridia and Microconidia in stem & then to leaves is common reaching in xylem and phloem and other tissues of the both (stem & leaf) and nomenclature of the pathogen may be given as *Fusarium oxysporum* var. *rosae*,

Acknowledgement

I have great regard to Dr. A.K. Mishra, Principal Scientist, Division of crop protection, Central Institute for Sub-tropical Horticulture, Rehmankhera, Kakori, Lucknow, U.P, India & Dr. R.K. Sharma, Senior Scientist (Mushroom), Division of Plant Pathology, I.A.R.I, New Delhi, India for their kind encouragement to work always in the field of fungi.

References

1. Devi , P.H.S et.al (2010) Effect of mulching on wiltincidence and yield of Pea (*Pisum sativum L.*) Indian Phytopath. 63(3) p. 348-349. Dorjey, S et. al (2017) Evolution of *Psuedomonas fluorescens* isolates for the management of *Fusarium oxysporum* f. sp. *Lycopesici* and *Rhizoctonia solani* Causing welt complex in Tomato. Indian Phytopath. 70 (1) p 127-130.
Dube, H.C (1983) An introduction to Fungi. Vikash Publishing House Pvt. Ltd., 5-Ansari Road, New Delhi-110002.

Dubey, S. C (2016) Race profiling genetic diversity, diagnostics & management of *F. oxysporum* f.sp.*ciceris* causing welt in chick Pea. Indian Phytopath. 69 (3) p210-217.
Dubey, S.C. et. al (2017) Evalution of chickpea genotypes against *Fusarium* wilt for resistance sources. Indian Phytopath. 70(2) p. 254 - 255.

6. Ingold, C.T (1971) The Biology of Fungi. The English Language Book Society & Hutchinson Educational Ltd., 3 Fitzroy Square, London WI.

7. Kaur, H et.al (2016) Morphological & Pathological characterization of *Fusarium verticilliodes* from different maize growing area of Punjab. Indian Phytopath. 69 (2) p 190–194.

8. Kaur, A et.all (2019) Effect of Abscisic Acid & Salicyclic Acid on growth and phenolic parameters under *Fusarum fujikuroi* infection in rice seedlings. Indian Phytopath. 72(2) p253-260.

9. Khan, P et.al (2017) Management of lettuce root caused by, *Fusarium oxysporum* f. *sp. lactucae* in hydroponically grown Crop using microbial consortia. Indian Phytopath 70(2) p232–237

10. Kotari, P et.al (2018) Molecular Investigations of gene expression analysis in two constracting genetypes of banana during Fusarium wilt (Foc1) infection. Indian Phytopath. 71(3) p.445-452.

11. Mehrotra, R.S (1980) Plant Pathology Tata Mc. Graw-Hill Publishing Co. Ltd., 12/4 Asaf Ali Road, New Delhi-110002.

12. Mishra, K.K (2008) Date Palm without date due to *Fusarium oxysporiem* var. *toddy* in Eastern zone of Nepal, 95th Indian Science Congress, Jan 3-7, 2008, Vishakhapatnum p.87.

13. Mishra, K.K (2012) *Fusarium oxysporum* on *Mimosa pudica* in eastern zone of Nepal at Biratnagar. 65th Ann. Meet. Indian Institute of Horticulture Research, Hessaraghatta Lake Post, Banglore, 4-6 Dec. 2012.

14. Mishra, K.K (2012) wilting in Mangifera indica having *Fusarium oxysporum* in Eastern zone of Nepal at Biratnagar. 65th Ann. Meet. IIHR, Hessaraghatta Lake Post, Banglore, 4-6 Dec, 2012.

15. Mishra, S & Dhar, V (2010) Identification of Broad base & stable sources of Resistance to Fusarium wilt in Pigeon Pea. Indian Phytopath. 63(2), p 165-167.

16. Mukhopadhyay, R et. al (2017) Moisture & Nitrogen stress induces severity of wilt nematode complex of lentil grown in rice fallow in an inceptisol. Indian Phytopath. 70 (1) p 109-113.

17. Murumkar, D.R (2010) variability among isolates of *Fusarium oxyspor<u>um</u>* f. sp. *Carthami* from Madhya Press. Indian Phytopath. 63(4) p 446-448.

18. Mushrif, S.K & Khulbe, D (2010) Genetic variability based on isozyme polymorphic among *Fusarium oxysporum* f.sp. *ciceri*-isolates inciting wilt in Chicken Pea. Indian Phytopath. 63(4) p 427-429.

19. Nazia, B & Sharda, M.S (2017) Enhancement of growth, defense response in brinjal and inhibitory potential of Salicyclic acid against *Fusarium oxysporum* sp. *melongenae*. Indian Phytopath. 70 (3) p 391-393.

20. Ramappa, H.K & Gowri, R (2016) Evaluation of Pigeon Pea genotypes against Fusarium wilt under artificial inoculation conditions. Indian Phytopath. 69(4) p381-385.

21. Roy, H & Sharma, S.D (2010) combination of soil solarization, vascular-Arbuscular Mycorrhiza & Azoto Bactor chrococcum for the management of seedling wilt of Citrus. Indian Phytopath. 63(3) p 282-285.

22. Raj, H et al (2016) Integrated management of Carnation wilt caused by *Fusarium oxysporum* f sp. *dianthi* using soil solarization and Soil amendments Indian Phytopath 69(4) p. 396-399.

23. Saharan, M.S (2010) Detection of genetic variation among *Fusarium spp.* collected from wheat growing regions of India. Indian Phytopath. 63(2) p 149-153.

24. Singh, P.K et. al (2010) Interactions of vesicular Arbuscular Mycorrhizal fungi with Fusarium wilt and growth of Tomato. Indian Phytopath. 63 (1) p 30-34.

25. Singh, R.S (1968) Plant diseases. Oxford & IBH Publishing Co., 17, Park street, Calcutta-16.

26. Singh, R.S (1980) Introduction to Principles of Plant Pathology oxford & IBH Publishing Co, 66 Janpath, New Delhi-110001.

27. Synder, W.C & Hansen, H.N (1940) The species concept in Fusarium. Amer. J. Bot. 27 p 64-67.

28. ----- (1941) The species concept in Fusarium with reference to Section Mortiella. Amer. J. Bot. 28 p 738-742.

29. ----- (1945) The species concept in Fusarium with reference to discolor & other species. Amer. J. Bot. 32 p 657-666.

30. Synder, W.C (1965) Current status of Taxonomy in *Fusarium species* and their Perfect stages. Phytopathology. 22 (8) p 833-837.

31. Wavare, S.H et. al (2017) Antifungal efficacy of floral extracts, biocontrol agents and fungicides against *Fusarium oxysportnum* f. sp. *Ciceri*. Indian phytopath. 70(2) p.191-199. 32. Wollen weber, H.W & Reinking. O.A (1938) DLE FUSARIEN. PAUL PAREY, BERLIN.

55042